

06-23-00

A

PTO/SB/05 (4/98)

09/30/2000. OMB 0651-0032

Patent and Trademark Office: U.S. DEPARTMENT OF COMMERCE

+

Under the Paperwork Reduction Act of 1995, no persons are required to respond to a collection of information unless it displays a valid OMB control number.

<b>UTILITY</b> <b>PATENT APPLICATION</b> <b>TRANSMITTAL</b> (Only for new nonprovisional application under 37 C.F.R. § 1.53(b))	Attorney Docket No.	99-2085
	First Inventor or Application Identifier	A. HOPPER
	Title	Compositions and Methods for the Quantification
	Express Mail Label No.	EL607418861US

<b>APPLICATION ELEMENTS</b> See MPEP chapter 600 concerning utility patent application contents	<b>ADDRESS TO</b> Assistant Commissioner for Patents Box Patent Application Washington DC 20231
--	--

1. <input checked="" type="checkbox"/> * Fee Transmittal Form (e.g., PTO/SB/17) (Submit an original and a duplicate for fee processing)	5. <input type="checkbox"/> Microfiche Computer Program (Appendix)
2. <input checked="" type="checkbox"/> Specification [Total Pages <b>43</b> ] (preferred arrangement set forth below)	6. Nucleotide and/or Amino Acid Sequence submission (if applicable, all necessary)
- Descriptive title of the invention - Cross References to Related Applications - Statement Regarding Fed sponsored R & D - Reference to Microfiche Appendix - Background of the invention - Brief Summary of the invention - Brief Description of the Drawings (if filed) - Detailed Description - Claim(s) - Abstract of the Disclosure	a. <input type="checkbox"/> Computer Readable Copy b. <input type="checkbox"/> Paper Copy (identical to computer copy) c. <input type="checkbox"/> Statement verifying identity of above copies
3. <input checked="" type="checkbox"/> Drawing(s) (35 U.S.C. 113) [Total Sheets <b>6</b> ]	<b>ACCOMPANYING APPLICATION PARTS</b>
4. Oath or Declaration [Total Sheets <b>4</b> ]	7. <input type="checkbox"/> Assignment Papers (cover sheet & document(s))
a. <input checked="" type="checkbox"/> Newly executed (original copy)	8. <input type="checkbox"/> 37 C.F.R. §3.73(b) Statement (when there is an assignee) <input type="checkbox"/> Power of Attorney
b. <input type="checkbox"/> Copy from a prior application (37 C.F.R. §1.63(d)) (for continuation/divisional with Box 16 completed)	9. <input type="checkbox"/> English Translation Document (if applicable)
i. <input type="checkbox"/> DELETION OF INVENTOR(S) Signed statement attached deleting inventor(s) named in the prior application, see 37 C.F.R. §§1.63(d)(2) and 1.33(b)	10. <input type="checkbox"/> Information Disclosure Statement (IDS)/PTO-1449 <input type="checkbox"/> Copies of IDS Citations
* NOTE FOR ITEMS 1 & 13: IN ORDER TO BE ENTITLED TO PAY SMALL ENTITY FEES, A SMALL ENTITY STATEMENT IS REQUIRED (37 C.F.R. §1.27), EXCEPT IF ONE FILED IN A PRIOR APPLICATION IS RELIED UPON (37 C.F.R. §1.28)	11. <input type="checkbox"/> Preliminary Amendment
	12. <input checked="" type="checkbox"/> Return Receipt Postcard (MPEP 503) (Should be specifically itemized)
	13. <input checked="" type="checkbox"/> * Small Entity Statement(s) <input type="checkbox"/> Statement filed in prior application, Status still proper and desired (PTO/SB/09-12)
	14. <input type="checkbox"/> Certified Copy of Priority Document(s) (if foreign priority is claimed)
	15. <input type="checkbox"/> Other:

If a CONTINUING APPLICATION, check appropriate box, and supply the requisite information below in a preliminary amendment

☐ Continuation ☐ Divisional ☐ Continuation-in-part of prior application No.: Claiming priority to 60/141,516 filed 6/23/99

Prior application information: Examiner \_\_\_\_\_ Group / Art Unit: \_\_\_\_\_ & 60/199,699 (4/26/00)

**For CONTINUATION or DIVISIONAL APPS only:** The entire disclosure of the prior application, from which an oath or declaration is supplied under Box 4b, is considered a part of the disclosure of the accompanying continuation or divisional application and is hereby incorporated by reference. The incorporation can only be relied upon when a portion has been inadvertently omitted from the submitted application parts.

17. CORRESPONDENCE ADDRESS					
<input type="checkbox"/> Customer Number or Bar Code Label			or <input type="checkbox"/> Correspondence address below		
(Insert Customer No. or Attach Bar code label here)					
Name	Thomas J. Monahan				
	Intellectual Property Office				
Address	The Pennsylvania State University				
	113 Technology Center				
City	University Park	State	PA	Zip Code	16802-7000
Country	United States	Telephone	(814) 865-6279	Fax	(814) 865-3591
Name (Print / Type)	Thomas J. Monahan		Registration No. (Attorney/Agent)		29,835
Signature				Date:	6/22/00

Burden Hour Statement: This form is estimated to take 0.2 hours to complete. Time will vary depending upon the needs of the individual case. Any comments on the amount of time you are required to complete this form should be sent to the Chief Information Officer, Patent and Trademark Office, Washington, DC 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SENT TO: Assistant Commissioner for Patents, Box Patent Application, Washington, DC 20231

 of Sterol Biosynthetic  
 Flux

Practitioner's Docket No.: 99-2085

**PATENT**

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

In re application of: A. HOPPER, et al.

Serial No.:

Group No.:

Filed: 06/22/00

Examiner:

For: COMPOSITIONS AND METHODS FOR THE QUANTIFICATION OF STEROL  
BIOTHYTHETIC FLUX

**Assistant Commissioner for Patents  
Washington D.C. 20231**

**EXPRESS MAIL CERTIFICATE**

"Express Mail" label number: EL607418861US

Date of Deposit: June 22, 2000

I hereby state that the following *attached* paper or fee:

- 1 Fee Transmittal Form (in duplicate)
- 1 Utility Patent Application
- 1 Small Entity Statement
- ☒ executed Declarations
- 43 pages of Specifications
- 6 sheets of Drawings
- 1 Return Receipt Postcard

Is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10, on the date indicated above and is addressed to the Assistant Commissioner for Patents, Washington, D.C. 20231.

Ruth A. Harpster

Typed or printed name of person mailing paper or fee

Ruth A. Harpster

Signature of person mailing paper or fee

**NOTE:** *The label number need not be placed on each page. It should, however, be placed on the first page of each separate document, such as, a new application, amendment, assignment, and transmittal letter for a fee, along with the certificate of mailing by "Express Mail". Although the label number may be on checks, such a practice is not required. In order not to deface formal drawings, it is suggested that the label number be placed on the back of each formal drawing or the drawings be accompanied by a set of informal drawings on which the label is placed.*

(Express Mail Certificate [8-3])

Under the Paperwork Reduction Act of 1995, no persons are required to respond to a collection of information unless it displays a valid OMB control number.

**STATEMENT CLAIMING SMALL ENTITY STATUS  
(37 CFR 1.9(f) & 1.27(d))--NONPROFIT ORGANIZATION**

Docket Number (Optional)  
99-2085

Applicant, Patentee, or Identifier: A. HOPPER, et al.

Application or Patent No.: \_\_\_\_\_

Filed or Issued: June 22, 2000

Title: COMPOSITIONS AND METHODS FOR THE QUANTIFICATION OF STEROL BIOSYNTHETIC FLUX

I hereby state that I am an official empowered to act on behalf of the nonprofit organization identified below:

NAME OF NONPROFIT ORGANIZATION The Penn State Research Foundation

ADDRESS OF NONPROFIT ORGANIZATION 304 Old Main  
University Park, PA 16802

**TYPE OF NONPROFIT ORGANIZATION:**

- ☐ UNIVERSITY OR OTHER INSTITUTION OF HIGHER EDUCATION
- ☒ TAX EXEMPT UNDER INTERNAL REVENUE SERVICE CODE (26 U.S.C. 501(a) and 501(c)(3))
- ☐ NONPROFIT SCIENTIFIC OR EDUCATIONAL UNDER STATUTE OF STATE OF THE UNITED STATES OF AMERICA  
(NAME OF STATE \_\_\_\_\_)  
(CITATION OF STATUTE \_\_\_\_\_)
- ☐ WOULD QUALIFY AS TAX EXEMPT UNDER INTERNAL REVENUE SERVICE CODE (26 U.S.C. 501(a) and 501(c)(3))  
IF LOCATED IN THE UNITED STATES OF AMERICA
- ☐ WOULD QUALIFY AS NONPROFIT SCIENTIFIC OR EDUCATIONAL UNDER STATUTE OF STATE OF THE UNITED STATES OF AMERICA IF LOCATED IN THE UNITED STATES OF AMERICA  
(NAME OF STATE \_\_\_\_\_)  
(CITATION OF STATUTE \_\_\_\_\_)

I hereby state that the nonprofit organization identified above qualifies as a nonprofit organization as defined in 37 CFR 1.9(e) for purposes of paying reduced fees to the United States Patent and Trademark Office regarding the invention described in:

- ☒ the specification filed herewith with title as listed above.
- ☐ the application identified above.
- ☐ the patent identified above.

I hereby state that rights under contract or law have been conveyed to and remain with the nonprofit organization regarding the above identified invention. If the rights held by the nonprofit organization are not exclusive, each individual, concern, or organization having rights in the invention must file separate statements as to their status as small entities and that no rights to the invention are held by any person, other than the inventor, who would not qualify as an independent inventor under 37CFR 1.9(c) if that person made the invention, or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

Each person, concern, or organization having any rights in the invention is listed below:

- ☐ no such person, concern, or organization exists.
- ☐ each such person, concern, or organization is listed below.

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

NAME OF PERSON SIGNING Thomas J. Monahan (Reg. No. 29,835)

TITLE IN ORGANIZATION OF PERSON SIGNING Director, Intellectual Property Office

ADDRESS OF PERSON SIGNING 113 Technology Center, University Park, PA 16802-7000

SIGNATURE [Signature] DATE June 22, 2000

## COMPOSITIONS AND METHODS FOR THE QUANTIFICATION OF STEROL BIOSYNTHETIC FLUX

This invention was made in part with government support under grants  
5 MCB9506810 and MCB9828216 from the National Science Foundation. The  
government has certain rights in the invention.

### FIELD OF THE INVENTION

This invention generally relates to a novel assay for the screening of compounds  
10 that are agonistic or antagonistic to the mevalonate pathway and sterol and cholesterol  
synthesis. In selected embodiments this assay incorporates colorimetric, growth, and  
immunological methods for high throughput screening of compounds.

### BACKGROUND

A finely tuned mechanism regulates the biosynthesis of mevalonate, the precursor  
5 of isoprenoid groups that are incorporated into more than a dozen classes of end products.  
These include: sterols, especially cholesterol, involved in membrane structure; haem A  
and ubiquinone, involved with electron transport; dolichol, required for glycoprotein  
synthesis; isopentenyladenine, present in some transfer RNAs; and intercellular  
20 messengers, such as cytokines in plants, farnesylated mating factors in fungi, juvenile  
hormones in insects and steroid hormones in animals. Interest in the regulatory  
importance of mevalonate was heightened by the discovery that growth-regulating p21<sup>ras</sup>  
proteins (encoded by *ras* proto-oncogenes and oncogenes) and nuclear envelope proteins,  
are covalently attached to farnesyl residues. These farnesyl residues, in turn, anchor said  
25 proteins to cell membranes. Inhibition of mevalonate synthesis prevents farnesylation of  
these proteins and blocks cell growth. To ensure constant production of the multiple  
isoprenoid compounds at all stages of growth, cells must precisely regulate mevalonate  
synthesis while avoiding over accumulation of potentially toxic products such as  
cholesterol. (Goldstein and Brown, "Regulation of the mevalonate pathway" *Nature*  
30 343:425-430, 1990).

The ability to regulate flux through the mevalonate pathway (Figure 1) is of  
medical importance because inhibitors of this pathway have been used to treat  
hypercholesterolemia and, consequently, to diminish the risk of heart attack. (Endo, "The  
discovery and development of HMG-CoA reductase inhibitors" *J Lipid Res* 33:1569-  
35 1582, 1992). Alteration of the pathway also affects the function of oncogenes (Reviews:  
Gibbs and Oliff, "The potential of farnesyltransferase inhibitors as cancer

chemotherapeutics" *Annu Rev Pharmacol Toxicol.* 37:143-66, 1997. The mevalonate pathway is an important target for many areas of therapeutic research and application. For example, HMG-CoA reductase catalyzes the rate-limiting step of the mevalonate pathway (Voet and Voet, *Biochemistry* Wiley, New York, 1990); therefore, inhibitors of this reductase have been developed for administration to patients with hypercholesterolemia in an attempt to lower their blood cholesterol levels (Endo, *et al.*, "Oxygenated cholesterol as ligands for cytosolic-nuclear tumor promoter binding protein: yakkasteroids" *Biochem Biophys Res Commun* 194:1529-35, 1993). Data reveals that the "statin" class (*eg.*, compactin and lovastatin; see Figure 1) of reductase inhibitors are reasonably safe and somewhat effective in lowering total cholesterol levels and preventing the progression and reducing the occurrence of coronary disease events (Brown, *et al.*, "Regression of coronary artery disease as a result of intensive lipid-lowering therapy in men with high levels of apolipoprotein B" *N Engl J Med* 323:1289-98, 1990; Endo, *et al.*, "Beneficial effects of dietary intervention on serum lipid and apolipoprotein levels in obese children" *Am J Dis Child* 146:303-305, 1992; Nash, *et al.*, "Meeting national cholesterol education goals in clinical practice--a comparison of lovastatin and fluvastatin in primary prevention" *Am J Cardiol.* 78 (Suppl. 6A):26-31, 1996, but more progress needs to be made in the development of therapies that are more effective.

As an indication of the breadth of potential therapeutic effect regulators of the mevalonate pathway can have, the statins, in addition to regulating cholesterol levels, also stimulate nitric oxide production (Endres *et al.*, "Role of peroxynitrite and neuronal nitric oxide synthase in the activation of poly(ADP-ribose) synthetase in a murine model of cerebral ischemia-reperfusion" *Neurosci Lett.* 248:41-41, 1998; Laufs *et al.*, "Upregulation of endothelial nitric oxide synthase by HMG CoA reductase inhibitors" *Circulation* 97:1129-35, 1998), have antiproliferative affects on some types of cancer cells (Lee *et al.*, "Inhibition of the 3-Hydroxy-3-methylglutaryl-coenzyme A reductase pathway induces p53-independent transcriptional regulation of p21<sup>waf1/cip1</sup> in human prostate carcinoma cells" *J. Biol. Chem.* 273: 10618-10623, 1998) and have immunosuppressive affects. Zaragozic acid inhibits the enzyme activity of squalene synthase which is the first step of the pathway committed solely to sterol biosynthesis, but appears not to be currently used in the clinical setting. Zaragozic acid D3 inhibits farnesyl-protein transferase and, therefore, protein prenylation as well (Tanimoto *et al.*, "Inhibitory activity to protein prenylation and antifungal activity of zaragozic acid D3, a potent inhibitor of squalene synthase produced by the fungus, *Mollisia sp* SANK 10294" *J Antibiot (Tokyo)* 51:428-431, 1998).

A screen for compounds affecting various steps of the mevalonate pathway, therefore, could identify potential therapeutics for treatment of hypercholesterolemia and other pathological conditions associated with sterol metabolism in addition to compounds which may inhibit oncogene protein prenylation and farnesylation. Moreover, the availability of a screen for flux through the sterol pathway could be useful for predicting undesirable side effects of drugs designed to treat other illnesses. Furthermore, since the mevalonate pathway is common to most organisms, compounds that regulate the mevalonate pathway may have uses beyond medicine such as agriculture and pest control. Therefore, what is needed is an efficient, flexible, high-throughput assay to screen for agents that are agonistic or antagonistic to mevalonate pathway function.

### SUMMARY OF THE INVENTION

The present invention relates to an assay designed to detect flux in the mevalonate pathway. In one embodiment the assay is a plate based assay incorporating the yeast strain *Saccharomyces cerevisiae*. Although it is not intended that the present invention be limited to a specific mechanism, it is believed that the modification of tRNA by Mod5p is in competition with flux through the mevalonate pathway. This competition results from both Mod5p and Erg20p using the same substrate, dimethylallyl-PP (Figure 1). Mod5p catalyzes the transfer of an isopentenyl moiety to an adenosine generating i<sup>6</sup>A at position 37 of some tRNAs. This modification affects the function of the tRNA in translation and may be measured by monitoring nonsense suppression. As shown in Figures 2 & 5F, two yeast strains have been generated that possess limiting cytosolic levels of Mod5p. When there is increased flux through the mevalonate pathway, in one example, by overproduction of Erg20p (Figure 1), there is less i<sup>6</sup>A modification of tRNA (Figure 3). This decreases the proliferation of cells which grow on media that will support strains expressing normal levels of Erg20p. See, Figure 2. Such a highly sensitive assay can differentiate as little as a two-fold difference in the level of i<sup>6</sup>A modification of tRNA. (Benko, *et al.*, "Competition between a sterol biosynthetic enzyme and the tRNA modification in addition to changes in the protein synthesis machinery causes altered nonsense suppression" *PNAS* 97:61-66, 2000).

Increased cytosolic levels of Mod5p cause a different phenotype easily assayed by growth on media lacking lysine (Zoladek *et al.*, "Mutations altering the mitochondrial-cytoplasmic distribution of Mod5p implicate the actin cytoskeleton and mRNA 3' ends and/or protein synthesis in mitochondrial delivery" *Mol Cell Biol.* 15:6884-6894, 1995), thereby, providing a means for selecting reagents that decrease flux through the pathway. In one embodiment of the present invention, therefore, yeast growth on particular media

can provide an index for both increases and decreases in mevalonate pathway flux. The assays contemplated by the present invention are efficient, inexpensive and provide new methods for screening drugs that alter the clinically significant mevalonate pathway.

It is not intended that the present invention be limited to the identification of compounds for only to a specific therapeutic application. However, in selected embodiments, compounds that regulate the mevalonate pathway may be beneficial in: (1) screening for new drugs to treat hyperlipidemias and other disorders in sterol metabolism such as Addison's disease and Cushing's syndrome; (2) screening for drugs that inhibit the function of farnesylated and prenylated oncogene products; (3) monitoring other drugs for possible side effects in sterol metabolism; (4) identification of yeast mutants with altered sterol metabolism; (5) screening for agents that alter plant physiological processes such as, for example, photosynthesis, cell growth, respiration, architecture and defense against pathogen attack (e.g., antibiotics and antifungals).

The present invention relates to a flexible, high throughput screen for agents that are agonistic or antagonistic to mevalonate pathway function. The present invention is not limited to any particular high throughput assay. Many high throughput assays are contemplated by the present invention. For example, in one embodiment, the present invention contemplates visually scoring plates. In another embodiment, the present invention contemplates culturing cells in microtiter plates, performing the assay, lysing the cells and generating a readout of said cell lysates via a spectrophotometer. In another embodiment, the present invention contemplates reading the cells in a flow cytometer to detect changes in cell color and in cell growth. In another embodiment, the present invention contemplates measuring (in one example by scintillation) radiation generated by H<sup>3</sup>-tritium as an index to determine cell growth. In preferred embodiments, the assay may be performed manually or with the use of automation and robotics for any or all steps in the aforementioned procedures.

The present invention relates to yeast strains engineered to provide detectable read-outs for compounds that are agonistic or antagonistic to the mevalonate pathway. In a preferred embodiment, the yeast strains are ALB1 (genotype: *MAT $\alpha$  mod5-M2 SUP7 ade2-1 can1-100 leu2-3, -112 lys1-1 lys2-1 trp1 ura3-1*), ALB8 (genotype: *MAT $\alpha$  SUP7 can1-100 ade2-1 leu2-3, -112 lys1-1 lys2-1 trp1 mod5::TRP1 ura3-1::MOD5*) and MT8-1D or MD14A with YCf mod5-m2KR6 plasmid for a lysine based assay.

It is not intended that the present invention be limited to the screening of any particular compound or class of compounds. Proteins, lipids, carbohydrates, glycoproteins, lipoproteins, synthetic compounds, compounds contained in combinatorial

libraries or compounds and agents already being used as therapeutics may be screened by the present assay. Moreover, the screening of known therapeutics according to the present invention will reveal, heretofore, unknown side effects associated with the modulation of the mevalonate pathway.

5 It is not intended the present invention be limited to any particular protocol to quantitate or qualitate the assay. In selected embodiments, for example, the assay output may be measured visually, colorimetrically, fluorescently, by immunoblot, by radio-immunoassay or by HPLC. In a one embodiment, anti-  $i^6A$  antibody is used to detect  $i^6A$  production in cells treated with a compound. In a preferred embodiment, a Western blot  
10 is used to detect the presence of  $i^6A$ . Quantitation is not limited to any particular method. Quantitation may be made by densitometer, chemiluminescence, and radiography.

The present invention contemplates a composition comprising yeast with the relevant genotype of: *SUP7 ade2-1 can1-100 leu2-3 mod5-M2* and designated ALB1. The present invention further contemplates a composition wherein the yeast ALB1 is a strain of *Saccharomyces cerevisiae*. Still further, the present invention contemplates a composition comprising yeast with the relevant genotype of: *SUP7 can1-100 ade2-1 leu2-3 mod5::TRP1 ura3-1::MOD5* and designated ALB8. Even still further, the present invention contemplates a composition wherein the yeast ALB8 is a strain of *Saccharomyces cerevisiae*. Even further still, the present invention contemplates a composition comprising the yeast strain ALB1 wherein the genotype further comprises: *MAT $\alpha$  mod5-M2 SUP7 ade2-1 can1-100 leu2-3, -112 lys1-1 lys2-1 trp1 ura3-1*. Even  
15 still further, the present invention contemplates a composition wherein the yeast ALB1 is a strain of *Saccharomyces cerevisiae*. Even further still, the present invention contemplates a composition comprising yeast with the relevant genotype of: *MAT $\alpha$ SUP7 can1-100 ade2-1 leu2-3, -112 lys1-1 lys2-1 trp1 mod5::TRP1 ura3-1::MOD5* and  
20 designated ALB8. Even further still, the present invention contemplates a composition of claim 7 wherein the yeast is *Saccharomyces cerevisiae*.

The present invention contemplates a method, comprising: a) providing: i) a test compound, ii) a growth media formulated to allow scoring of nonsense suppression in  
25 yeast, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p, or its homolog, as



compared to said wild type yeast cells, and wherein said modified yeast cells comprise a gene with a nonsense mutation and a suppressor tRNA gene coding for a tRNA modified with isopentenyl adenosine by Mod5 or its homolog; b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and c) measuring for growth of said treated portion.

The present invention also contemplates a measuring of step which comprises examining the color of said yeast cells of said treated portion.

Additionally, the present invention contemplates a measuring of step which comprises comparing said treated portion with said untreated portion, wherein said untreated portion is exposed to said growth media in the absence of said test compound.

The present invention contemplates a method, comprising: a) providing: i) a test compound, ii) a growth media lacking adenine, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise an *ADE* gene having a nonsense mutation and a gene coding for a nonsense suppressor tRNA; b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and c) measuring for growth of said treated portion.

The present invention also contemplates a method, comprising: a) providing: i) a test compound, ii) a growth media lacking adenine, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise an *ADE* gene having a nonsense mutation and a *SUP7* gene coding for a tRNA; b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and c) measuring for growth of said treated portion.

The present invention also contemplates a method, comprising: a) providing: i) a test compound, ii) a growth media lacking arginine and comprising a canavanine salt, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise a *CAN1* gene having a nonsense mutation

and a gene coding for a nonsense suppressor tRNA; b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and c) measuring for growth of said treated portion.

The present invention also contemplates a method, comprising: a) providing: i) a test compound, ii) a growth media lacking arginine and comprising a canavanine salt, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise a *CAN1* gene having a nonsense mutation and a *SUP7* gene coding for a tRNA; b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and c) measuring for growth of said treated portion.

Additionally, the present invention contemplates a method wherein said gene coding for said nonsense suppressor tRNA is selected from the group consisting of *SUP7* and *SUP11*.

The present invention further contemplates a method comprising: a) providing i) one or more compounds, ii) a first yeast cell line designated ALB1; ii) a second yeast cell line designated ALB8; b) contacting a portion of said cells from i) said first yeast cell line and ii) said second yeast cell line, with said one or more said compounds, so as to create treated portions and untreated portions of cells; and, c) comparing said treated cells with said untreated cells. Even further, the present invention contemplates that the method of comparison of said treated or untreated cells may be by color and cell growth (*i.e.*, the amount of cell division).

The present invention further contemplates a method comprising: a) providing i) one or more compounds and ii) a yeast cell line selected from a group consisting of yeast strains designated ALB1 and ALB8; b) contacting a portion of said cells from said yeast cell line with said one or more said compounds, so as to create treated portions and untreated portions of cells; and, c) i) comparing said treated cells with said untreated cells. The present invention further contemplates the comparison of treated and untreated cells by color and by cell growth.

## DESCRIPTION OF THE FIGURES

Figure 1 shows the mevalonate pathway.

Figure 2 shows the level of isopentenylated tRNA found in ALB1 over-expressing *ERG20* is substantially reduced.

A. Low molecular weight RNA was prepared from ALB1 (*mod5-M2*) with each of the candidate genes or vector alone, ALB8 (*MOD5*) or MD14A (*mod5-1*). The RNAs were resolved on polyacrylamide gels, transferred to membranes and probed with anti-isopentenyl adenosine antibody (upper panel) or radiolabeled oligonucleotide complementary to mature tRNA<sup>Tyr</sup> (lower panel).

B. The levels of isopentenyl adenosine tRNA found in ALB1 with each of the candidate genes or vector only or in the strain ALB8 or MD14A were assessed by densitometric analysis of two immunoblots and expressed as a fraction of the level found in the "vector" control. (A) membrane 1 values; (B) membrane 2 values; (C) average values.

Figure 3 presents a model of competition between i<sup>6</sup>A modification of tRNA and sterol biosynthesis.

Figure 4 presents selected mutations introduced into the *MOD5* gene and oligonucleotides. See, Gillman *et al.*, "*MOD5* translation initiation sites determine N6-isopentenyladenosine modification of mitochondrial and cytoplasmic tRNA", "*Mol Cell Biol* 11:2382-2390.

Figure 5 presents two yeast strains that present limited cytosolic levels of Mod5p. Cells with the *mod5-M2KR6* allele (T8-ID with YCfmod5-M2KR6 as projected in Fig. 5) have a very small cytosolic pool of Mod5p and the cells are unable to grow in the absence of lysine.

Figure 6 presents a restriction map of YEpMOD5(7.0) and subclones YEpMOD5(1.8) and YEpMOD5(1.9). Numbers in parentheses indicate size of insert. See, Dihanich, *et al.*, "Isolation and characterization of *MOD5*, a gene required for isopentenylation of cytoplasmic and mitochondrial tRNAs of *Saccharomyces cerevisiae*" "*Mol Cell Biol* 7:177-184, 1987).

## DEFINITIONS

To facilitate understanding of the invention, a number of terms are defined below.

"Genotype" shall be defined as the genetic makeup of an organism as encoded in its DNA.

"Relevant genotype" shall be defined as specific genes that, if expressed, have impact on the investigation under way.

A "nonsense suppressor tRNA" is meant to indicate one of several known suppressor tRNAs. For example, *SUP7-1* codes for an altered tRNA that is one of several efficient tyrosine-inserting UAA suppressors. (H. Laten *et al.*, Nucleic Acids Res. 5:4329 (1978)). Indeed, *SUP7* (and *SUP 11* for that matter) can suppress the *lys2-1* nonsense allele (as well as other nonsense mutations). The *LYS2* gene encodes an enzyme involved in the biosynthesis of lysine. Mutations in this gene prevent growth on media lacking exogenous lysine.

As used herein, the term "scoring of nonsense suppression in yeast" refers to determining the level of suppression due to translation through the nonsense codon in mRNA relative to a control sample wherein said control sample comprises reduced or absent translation through the nonsense codon in mRNA because of differences in suppressor tRNA.

As referred to herein, cells with the mod5-M2 allele (strain *ALB1* projected in Fig. 2) contains a mutation at codon 12 preventing the initiation of Mod5p-II translation (and allow only Mod5p-I production) have about 50% of the *SUP7* pool modified resulting in intermediate levels of suppression. These cells show intermediate growth on canavanine and on media lacking adenine.

As referred to herein, cells with the mod5-M2KR6 allele (T8-ID with Ycfmod5-M2KR6 as projected in Fig. 5) have a very small cytosolic pool of Mod5p and the cells are unable to grow in the absence of lysine.

Yeast strain "ALB1" is defined as a substantially pure population of yeast with the genotype of *MAT $\alpha$  mod5-M2 SUP7 ade2-1 can1-100 leu2-3, -112 lys1-1 lys2-1 trp1 ura3-1* and the relevant genotype of: *SUP7 ade2-1 can1-100 leu2-3-112 mod5-M2*. The yeast may be of the species *Saccharomyces cerevisiae*.

Yeast strain "ALB8" shall be defined as a substantially pure population of yeast with the genotype of: *MAT $\alpha$  SUP7 can1-100 ade2-1 leu2-3, -112 lys1-1 lys2-1 trp1 mod5::TRP1 ura3-1::MOD5*. The yeast may be of the species *Saccharomyces cerevisiae*.

Yeast strain "T8-1D" shall be defined as a substantially pure population of yeast with the genotype of: *MAT $\alpha$  SUP11 ade2-1 leu2-3, -112 mod5-1 lys2-1 his4-519 ura 3-1*. The yeast may be of the species *Saccharomyces cerevisiae*.

As used herein, *CAN1* refers to a sequence encoding an arginine permease that allows the uptake of the arginine analog canavanine. Canavanine interferes with the process of translation and cells cannot grow in its presence. Therefore, cells with wild-type *CAN1* are sensitive to canavanine, but cells with the mutant *can1-100*, that does not encode a properly functioning permease, are resistant to canavanine and can grow in its presence. See, Bun-Ichiro Ono, *et al.*, "Nonsense Mutations in the *can1* Locus of *Saccharomyces cerevisiae*", Journal of Bacteriology, June 1983, pp. 1476-1479.

As used herein, *ADE2* refers to a sequence which encodes an enzyme involved in the synthesis of adenine. Cells with *ade2-1* turn red in color and fail to grow on defined medium lacking adenine (Ade), whereas cells producing functional Ade2p can grow on such medium (Ade+) and are white. Cells with the *ade2-1* allele and sufficient i6A-modified suppressor tRNA can grow in the absence of exogenous adenine and generate white colonies on rich medium, whereas cells with insufficient i6A-modified tRNA are unable to grow in the absence of exogenous adenine and generate red colonies on rich medium. Cells with intermediate levels of i6A modified tRNA have intermediate phenotypes in colony color and intermediate rates of growth in the absence of exogenous adenine.

As used herein a, "substantially deficient adenine growth media" refers to a culture media that, in one embodiment, has less than 20mg /L of adenine while in a preferred embodiment has less than 5mg/L of adenine.

"Wild type" shall be defined as the genomic makeup of an organism (the genotype) before modifications have been made. In other words, it is the parent strain of the organism.

The term "binding interaction" when used in relation to RNA shall be defined as the ability of two or more proteins to bind to each other (*e.g.*, to produce an aggregate). The present invention makes no limit on the stringency of the binding interaction so long as the interaction can be detected by methods known to those practiced in the art (*e.g.*, by Western blot, coimmunoprecipitation, spectrophotometry, colorimetric assay, etc.).

The term "homology" when used in relation to proteins refers to a degree of similarity. There may be partial homology or complete homology (*i.e.*, identity). A partially similar sequence is one that may partially inhibit a similar sequence from performing its function (*e.g.*, enzymatic, binding, etc) *in vivo* or *in vitro* and is referred to using the functional term "substantially homologous." The inhibition function of the

substantially similar sequence may be examined using an enzymatic assay, a binding assay or other assay designed to measure the particular function of the substantially similar protein. A substantially homologous proteins may compete for or interfere with a homolog and inhibit its function (*e.g.*, the binding or enzymatic function). This is not to say that conditions of low stringency are such that non-specific interaction is permitted; low stringency conditions require that the interaction be a specific (*i.e.*, selective) interaction.

Low stringency conditions when used in reference to nucleic acid hybridization comprise conditions equivalent to binding or hybridization at 42°C in a solution consisting of 5X SSPE (43.8 g/l NaCl, 6.9 g/l NaH<sub>2</sub>PO<sub>4</sub>•H<sub>2</sub>O and 1.85 g/l EDTA, pH adjusted to 7.4 with NaOH), 0.1% SDS, 5X Denhardt's reagent [50X Denhardt's contains per 500 ml: 5 g Ficoll (Type 400, Pharmacia), 5 g BSA (Fraction V; Sigma)] and 100 ug/ml denatured salmon sperm DNA followed by washing in a solution comprising 5X SSPE, 0.1% SDS at 42°C when a probe of about 500 nucleotides in length is employed.

High stringency conditions when used in reference to nucleic acid hybridization comprise conditions equivalent to binding or hybridization at 42°C in a solution consisting of 5X SSPE (43.8 g/l NaCl, 6.9 g/l NaH<sub>2</sub>PO<sub>4</sub>•H<sub>2</sub>O and 1.85 g/l EDTA, pH adjusted to 7.4 with NaOH), 0.5% SDS, 5X Denhardt's reagent and 100 µg/ml denatured salmon sperm DNA followed by washing in a solution comprising 0.1X SSPE, 1.0% SDS at 42°C when a probe of about 500 nucleotides in length is employed.

When used in reference to nucleic acid hybridization the art knows well that numerous equivalent conditions may be employed to comprise either low or high stringency conditions; factors such as the length and nature (DNA, RNA, base composition) of the probe and nature of the target (DNA, RNA, base composition, present in solution or immobilized, etc.) and the concentration of the salts and other components (*e.g.*, the presence or absence of formamide, dextran sulfate, polyethylene glycol) are considered and the hybridization solution may be varied to generate conditions of either low or high stringency hybridization different from, but equivalent to, the above listed conditions.

"Stringency" when used in reference to nucleic acid hybridization typically occurs in a range from about  $T_m - 5^\circ\text{C}$  (5°C below the  $T_m$  of the probe) to about 20°C to 25°C below  $T_m$ . As will be understood by those of skill in the art, a stringent hybridization can be used to identify or detect identical polynucleotide sequences or to identify or detect similar or related polynucleotide sequences. Under "stringent conditions" a nucleic acid sequence of interest will hybridize to its exact complement and closely related sequences.

As used herein, the term "purified" or "to purify" refers to the removal of contaminants from a sample. The present invention contemplates purified compositions (e.g., the ALB1 and ALB8 stains of *S. cerevisiae*).

As used herein, the term "substantially purified" refers to the removal of a significant portion of the contaminants of a sample to the extent that the substance of interest is recognizable by techniques known to those skilled in the art as the most abundant substance in the mixture.

As used herein the term "portion" when in reference to a protein (as in "a portion of a given protein") refers to fragments of that protein. The fragments may range in size from four amino acid residues to the entire amino acid sequence minus one amino acid. In one embodiment, the present invention contemplates "functional portions" of a protein. Such portions are "functional" if they contain a binding region (i.e., a region having affinity for another molecule) and such binding can take place (i.e., the binding region functions, albeit with perhaps lower affinity than that observed for the full-length protein). Such "functional portions" of the gene product are typically greater than 50 amino acids in length, and more typically greater than 100 amino acids in length. "Functional portions" may also be "conserved portions" of the protein. The alignment of the yeast and human gene products (described herein) permit one skilled in the art to select conserved portions of the protein (i.e., those portions in common between yeast and man) as well as unconserved portions (i.e., those portions unique to either yeast or man). The present invention contemplates conserved portions 20 amino acids in length or greater, and more typically greater than 50 amino acids in length.

As used herein the term "portion" when in reference to an oligonucleotide sequence (as in "a portion of a given sequence") refers to fragments of that sequence. The fragments may range in size from four base residues to the entire oligonucleotide sequence minus one base. More typically, such portions are 15 nucleotides in length or greater. Again, such portions may be conserved portions (see Figure 8). On the other hand, such portions may be unique portions of the gene.

"Compound" refers to any chemical entity, pharmaceutical, drug, and the like that can be used to treat or prevent a disease, illness, sickness, or disorder of bodily function. Compounds comprise both known and potential therapeutic compounds. A compound can be determined to have therapeutic potential by screening, e.g., using the screening methods of the present invention. A "known therapeutic compound" refers to a therapeutic compound that has been shown (e.g., through animal trials or prior experience with administration to humans) to be effective in such treatment or prevention.

"Antibody" shall be defined as a glycoprotein produced by B cells that binds with high specificity to the agent (usually, but not always, a peptide), or a structurally similar agent (*e.g.*,  $i^{6}A$ ), that generated its production (*e.g.*, the  $i^{6}A$  antibody). Antibodies may be produced by any of the known methodologies [Current Protocols in Immunology (1998) John Wiley and Sons, Inc., N.Y.] and may be either polyclonal or monoclonal.

"Antigen" shall be defined as a protein, glycoprotein, lipoprotein, lipid or other substance that is reactive with an antibody specific for a portion of the molecule.

The terms "immunoprecipitate", "immunoprecipitated", "immunoprecipitation" refer to the use of antibody to take an antigen out of solution by precipitation.

The term "affinity purification" refers to the use of an antibody to separate its antigen or a portion thereof from a mixture of other molecules because of affinity for the antigen.

"Immunofluorescence" is a staining technique used to identify, mark, label, visualize or make readily apparent by procedures known to those practiced in the art, where a ligand (usually an antibody) is bound to a receptor (usually an antigen) and such ligand, if an antibody, is conjugated to a fluorescent molecule, or the ligand is then bound by an antibody specific for the ligand, and said antibody is conjugated to a fluorescent molecule, where said fluorescent molecule can be visualized with the appropriate instrument (*e.g.*, a fluorescent microscope).

"Staining" shall be defined as any number of processes known to those in the field that are used to better visualize, distinguish or identify a specific component(s) and/or feature(s) of a cell or cells.

"RIA" or "radioimmunoassay" shall be defined as an assay wherein antibody is used to detect antigen and thereafter quantitated with radioisotopes.

"Immunoabsorbant" and "immunoaffinity" shall be defined as binding antigen with antibody reactive to said antigen. The antibody or the antigen may be bound to a solid or semi-solid substrate.

"Western blot" shall be defined as an assay comprising antigen that is detected by antibodies reactive to the antigen after the antigen has been at least partially separated from contaminants by, for example, electrophoresis.

"In operable combination", "in operable order" and "operably linked" as used herein refer to the linkage of nucleic acid sequences in such a manner that a nucleic acid molecule capable of directing the transcription of a given gene and/or the synthesis of a desired protein molecule is produced. The term also refers to the linkage of amino acid sequences in such a manner so that a functional protein is produced.



"Heterologous DNA" sequence refers to a nucleotide sequence which is not endogenous to the cell into which it is introduced. Heterologous DNA includes a nucleotide sequence which is ligated to, or is manipulated to become ligated to, a nucleic acid sequence to which it is not ligated in nature, or to which it is ligated at a different location in nature. Heterologous DNA also includes a nucleotide sequence which is naturally found in the cell into which it is introduced and which contains some modification relative to the naturally-occurring sequence.

"Morphology" shall be defined as the visual appearance of a cell or organism when viewed with the eye, a light microscope, a confocal microscope or an electronmicroscope, as appropriate. Morphology shall include, but is not limited to, cell shape, cell appearance and cell color.

"Patient" shall be defined as a human or other animal, such as a guinea pig or mouse and the like.

## GENERAL DESCRIPTION OF THE INVENTION

This invention generally relates to a novel assay for the screening of compounds that are agonistic or antagonistic to the mevalonate pathway and sterol and cholesterol synthesis. In selected embodiments this assay incorporates colorimetric, growth, biochemical (e.g., HPLC) and immunological methods for high through-put screening of compounds.

Baker's yeast *Saccharomyces cerevisiae* are single celled eukaryotes used extensively as model organisms. This model system provides for the facile manipulations of yeast genes, availability of the complete genomic sequence of baker's yeast, and identification of numerous yeast genes which directly correlate with known human disease genes. One fundamental discovery that has become clear from the use of this model system is that many physiologically significant regulatory pathways and proteins are highly conserved among all eukaryotes. Importantly, mammalian homologues of many yeast proteins are known to be important in regulating cell physiology, including the mevalonate pathway. Therefore, these model systems have been effectively utilized for the rapid identification and functional characterization of compounds that may be of use as therapeutics in higher eukaryotes. Thus yeast is well suited as a model system not only for the elucidation of basic cell biology but also for large-scale screening of compounds which specifically target cell growth.

## I. Assay Development

The *Saccharomyces cerevisiae* protein Mod5p catalyzes the addition of an isopentenyl group to adenosine ( $i^6A$ ) at position 37 of the anticodon loop of some tRNAs (Kline, *et al.*, "N6-(delta-2-isopentenyl) adenosine. Biosynthesis in transfer ribonucleic acid *in vitro*" *Biochemistry* 8:4361-4371, 1969; Bartz, *et al.*, "N6-(Delta 2-isopentenyl)adenosine: biosynthesis *in vitro* in transfer RNA by an enzyme purified from *Escherichia coli*" *Biochem Biophys Res Commun* 40:1481-1487, 1970; Rosenbaum and Gefter, "Delta 2 -isopentenylpyrophosphate: transfer ribonucleic acid 2-isopentenyltransferase from *Escherichia coli*. Purification and properties of the enzyme" *J Biol Chem* 247:5675-5680, 1972; McCloskey and Nishimura, *Acc. Chem. Res.* 10:403-410, 1977; Dihanich, *et al.*, "Isolation and characterization of *MOD5*, a gene required for isopentenylation of cytoplasmic and mitochondrial tRNAs of *Saccharomyces cerevisiae*" *Mol Cell Biol* 7:177-184, 1987). There are two isoforms of Mod5p, Mod5p-I and Mod5p-II, which differ in the site of their translation initiation codon and in their distribution in the cell. Mod5p-I is translated starting at codon one of the *MOD5* ORF and is localized to mitochondria and the cytoplasm. Mod5p-II, translated from codon twelve of the *MOD5* ORF, is located in the nucleus and the cytoplasm (Gillman, *et al.*, "MOD5 translation initiation sites determine N6-isopentenyladenosine modification of mitochondrial and cytoplasmic tRNA" *Mol Cell Biol* 11:2382-2390, 1991; Boguta, *et al.*, "Subcellular locations of *MOD5* proteins: mapping of sequences sufficient for targeting to mitochondria and demonstration that mitochondrial and nuclear isoforms commingle in the cytosol" *Mol Cell Biol* 14:2298-2306, 1994).

Over-expression of Erg20p causes a decrease in  $i^6A$  tRNA-mediated suppression in yeast and an approximately 70% decrease in  $i^6A$  modification of tRNA. These effects are most likely due to the loss of Mod5p substrate to Erg20p. The data demonstrate the dependence of tRNA processing upon changes in components of the sterol pathway. The data also indicate that, at least in yeast, Erg20p may also catalyze a rate-limiting step in sterol biosynthesis. Hence, the tRNA and the sterol biosynthetic pathways are in apparent competition and a delicate balance in protein levels is required to maintain proper functioning of translation and mevalonate metabolism.

The  $i^6A$  modification promotes the efficiency of *SUP7* or *SUP11* tRNA in cytosolic suppression of UAA nonsense mutations by the insertion of tyrosine (Laten, *et al.*, "Isopentenyladenosine deficient tRNA from an antisuppressor mutant of *Saccharomyces cerevisiae*" *Nucleic Acids Res* 5:4329-4342, 1978). Cells possessing only Mod5p-I have limiting cytosolic amounts of isozyme and changes in the subcellular distribution and/or the activity of this isozyme alter nonsense suppression. Hence genetic

screens/selections based on nonsense suppression can identify cells with altered cytosolic Mod5p-I activity (Zoladek, *et al.*, "Mutations altering the mitochondrial-cytoplasmic distribution of Mod5p implicate the actin cytoskeleton and mRNA 3' ends and/or protein synthesis in mitochondrial delivery" *Mol Cell Biol* 15:6884-6894, 1995).

Applicants employed the genetic strategy of using over expression to perturb a pathway (Rine, "Gene overexpression in studies of *Saccharomyces cerevisiae*" *Methods Enzymol* 194:239-251, 1991) and developed a protocol for the selection of cells with lower than normal levels of cytosolic Mod5p-I activity. Using this strategy Applicants were able to sample the entire yeast genome and identify genes that, when overexpressed, lead to lower than normal levels of cytosolic Mod5p-I activity.

As a result of that screen, Applicants have identified two categories of genes. The first category includes genes that affect nonsense suppression via alteration of the protein synthetic machinery. Applicants' studies suggest that the yeast gene product encoded by YDL219w may function in protein synthesis and that the translation elongation factor EF1- $\gamma$  may function in translational proofreading. *SAL6* and *ARC1*, with previously established effects on protein synthesis, were also recovered.

The second category includes *ERG20*, involved in sterol biosynthesis. The mevalonate pathway generates sterols, prenylated proteins, heme A, dolichol, ubiquinone and the substrate required for isopentenylation of tRNAs (Figure 1). Both Mod5p and Erg20p use the same substrate, dimethylallyl pyrophosphate (DMAPP). Recovery of *ERG20* coupled with subsequent biochemical assays demonstrating a reduction of i<sup>6</sup>A on tRNA is most easily explained by a model in which Mod5p and Erg20p compete for a limited pool of DMAPP. When more DMAPP is used to make sterols, less is available for modification of tRNA and a reduction in the efficiency of nonsense suppression results. Thus, Applicants have demonstrated that the tRNA biosynthetic pathway and the sterol biosynthetic pathway are in apparent competition for substrate and that Erg20p and Mod5p must be balanced to optimally maintain the protein synthetic machinery. A practical consequence is that it should be possible to adapt the selection the applicants developed to assess the effect of mutations and/or drugs that change the distribution of DMAPP between the sterol pathway and the tRNA biosynthetic pathway.

Applicants interest in the distribution of sorting isozymes led to the design of a screen for gene products that function in the subcellular distribution of Mod5p or affect its ability to modify cytosolic tRNAs. Applicants anticipated the discovery of genes affecting transcription of *MOD5* and/or tRNAs, nuclear export of tRNA, and the translation process, (in addition to those involved in protein distribution). Three gene products, *Sal6p*, *Tef4p* and *YDL219w*, may affect nonsense suppression via function in

protein translation and Arc1p could play a role in nuclear export of tRNA. No gene products affecting *MOD5* or tRNA transcription or Mod5p-I subcellular distribution were uncovered. Rather, *ERG20*, important to sterol biosynthesis, was discovered.

Sal6p/Ppq1p is a serine-threonine protein phosphatase very similar to mammalian phosphatase *PP1* (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994; Chen, *et al.*, "PPQ, a novel protein phosphatase containing a Ser<sup>+</sup> Asn-rich amino-terminal domain, is involved in the regulation of protein synthesis" *Eur J Biochem* 218:689-699, 1993). Multiple copies of *SAL6* cause antisuppression of nonsense mutations (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994) and the *sal6-1* allele acts as an allosuppressor (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994; Stansfield and Tuite, "Polypeptide chain termination in *Saccharomyces cerevisiae*" *Curr Genet* 25:385-395, 1994). Yeast cells containing a disrupted *PPQ1* gene exhibit a slowed translation rate and hypersensitivity to inhibitors of protein synthesis (Chen, *et al.*, "PPQ, a novel protein phosphatase containing a Ser + Asn-rich amino-terminal domain, is involved in the regulation of protein synthesis" *Eur J Biochem* 218:689-699, 1993). These findings suggest a role for Sal6p in the regulation of the fidelity of translation (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994; Chen, *et al.*, "PPQ, a novel protein phosphatase containing a Ser + Asn-rich amino-terminal domain, is involved in the regulation of protein synthesis" *Eur J Biochem* 218:689-699, 1993).

Tef4p is the gamma subunit of elongation factor 1 (EF-1). Eukaryotic EF-1 functions in delivering incoming tRNAs to the ribosome and is composed of at least three subunits. EF-1 $\alpha$  binds GTP and then the proper aminoacyl-tRNA and positions the aminoacyl tRNA in the ribosomal A site. Subsequently, GTP is hydrolyzed to GDP and EF-1 $\beta$  catalyzes the exchange of GDP for GTP to restore EF-1 $\alpha$  to its initial state (Kinzy, *et al.*, "Multiple genes encode the translation elongation factor EF-1 gamma in *Saccharomyces cerevisiae*" *Nucleic Acids Res* 22:2703-2707, 1994; Hinnebusch and Liebman, in "The molecular and cellular biology of the yeast *Saccharomyces*: Genomic dynamics, protein synthesis and energetics" eds. Broach, *et al.* [Cold Spring Harbor Lab Press, Plainview, NY] Vol. 1, pp. 627-735, 1991). The exact function of EF-1 $\delta$  has not been determined; however, in *Artemia salina* this subunit was observed to enhance the

function of EF-1 $\beta$  catalysis. In the same study EF1- $\gamma$  was found to interact with cell membranes and tubulin suggesting that it might mediate the association of the translational machinery with the cell framework (Janssen and Möller, "Elongation factor 1 beta gamma from *Artemia*. Purification and properties of its subunits" *Eur J Biochem* 171:119-129, 1988).

In bacteria the EF-1 counterpart functions in translational proofreading. Incorrectly inserted aminoacyl tRNAs can be removed prior to GTP hydrolysis and prior to the departure of EF-1 $\alpha$  from the ribosome (Hinnebusch and Liebman, in "The molecular and cellular biology of the yeast *Saccharomyces*: Genomic dynamics, protein synthesis and energetics" eds. Broach, *et al.* [Cold Spring Harbor Lab Press, Plainview, NY] Vol. 1, pp. 627-735, 1991). If the yeast EF-1 serves a similar function as the bacterial counterpart, it is unlikely that the  $\alpha$  subunit participates in this function as extra copies of the genes encoding EF-1 $\alpha$  decrease fidelity (*i.e.*, they enhance suppression of nonsense mutations; Song, *et al.*, "Elongation factor EF-1 alpha gene dosage alters translational fidelity in *Saccharomyces cerevisiae*" *Mol Cell Biol* 9:4571-4575, 1989). Applicants show that over-expression of Tef4p reduces nonsense suppression and therefore it is possible that it is the EF1- $\gamma$  subunit that functions in EF-1 proofreading.

ORF YDL219w is predicted to code for a 150 amino acid protein with no significant homology to any characterized protein. However two lines of evidence indicate that this protein may function in the translation process. First, the gene possesses an intron (*Saccharomyces* Genome Database, <http://genome-www.stanford.edu/cgi-bin/dbrun>). As introns are rare in yeast other than for approximately half of the genes encoding ribosomal proteins (Woolford and Warner, in "The molecular and cellular biology of the yeast *Saccharomyces*: Genomic dynamics, protein synthesis and energetics" eds. Broach, *et al.* [Cold Spring Harbor Lab Press, Plainview, NY] Vol. 1, pp. 587-626, 1991), the presence of the intron is suggestive of a role in translation. Second, Applicants show that over expression of YDL219w affects tRNA-mediated nonsense suppression.

Arc1p, or G4p1, was originally discovered as a quadruplex nucleic acid binding protein (Frantz and Gilbert, "A novel yeast gene product, G4p1, with a specific affinity for quadruplex nucleic acids" *J Biol Chem* 270:20692-20697, 1995). Subsequently, Arc1p was demonstrated to bind tRNA and to complex with methionyl and glutamyl tRNA synthetases (Simos, *et al.*, "The yeast protein Arc1p binds to tRNA and functions as a cofactor for the methionyl- and glutamyl-tRNA synthetases" *EMBO J* 15:5437-5448, 1996. Recent studies have shown that aminoacylation is important for tRNA nuclear export (Lund and Dahlberg, "Proofreading and aminoacylation of tRNAs before export

from the nucleus" *Science* 282:2082-2085, 1998; Sarkar, *et al.*, "Nuclear tRNA aminoacylation and its role in nuclear export of endogenous tRNAs in *Saccharomyces cerevisiae*" *PNAS* 96:14366-14371, 1999). Moreover, Simos *et al.* (Simos, *et al.*, "The yeast protein Arc1p binds to tRNA and functions as a cofactor for the methionyl- and glutamyl-tRNA synthetases" *EMBO J* 15:5437-5448, 1996) uncovered *ARC1* by synthetic interactions with Los1p, the yeast tRNA exportin (Arts, *et al.*, "Identification of a nuclear export receptor for tRNA" *Curr Biol* 8:305-314, 1998; Sarkar and Hopper, "tRNA nuclear export in *Saccharomyces cerevisiae*: *in situ* hybridization analysis" *Mol Biol Cell* 9:3041-3055, 1998). Thus, it is possible that over-expression of Arc1p interferes with the appropriate nucleus/cytosol distribution of tRNA. Alternatively, over-expressed Arc1p could interfere with the interaction of tRNA and ribosomes. Incubation of tRNA<sup>Met</sup> with large amounts of Arc1p *in vitro*, resulted in the creation of supershifted complexes in a gel mobility assay, indicative of higher-order complexes (Simos, *et al.*, "The yeast protein Arc1p binds to tRNA and functions as a cofactor for the methionyl- and glutamyl-tRNA synthetases" *EMBO J* 15:5437-5448, 1996). Perhaps, large quantities of cellular Arc1p complexes with tRNA, preventing the tRNA from functioning normally in translation.

It is contemplated that mutations may be introduced into the *MOD5* gene and oligonucleotides used in mutagenesis. Figure 4 presents the nucleotide sequence of the *MOD5* gene from two nucleotides 5' of the SnaBI site upstream of the open reading frame codon 14, followed by the sequence in the area of the first NsiI site. The DNA sequences of oligonucleotides M1 and M2 are shown below the corresponding wild-type (WT) sequence. Restriction endoculease recognition sites are underlined and indicated above the sites. Encoded amino acids are in three-letter code below codons. The first two methionine codons are indicated by capital letters. The M2 oligonucleotide encodes the only amino acid change in *MOD5* (Met to Ile); all other changes become part of the 5' noncoding portion of the gene, Arrows show positions of 5' ends of *MOD5* transcripts.

The mevalonate pathway generates sterols, prenylated proteins, heme A, dolichol, ubiquinone and isopentenyl tRNAs. Inhibitors of HMG-CoA reductase, catalyzing a rate-limiting step of the pathway, have been developed for administration to patients with hypercholesterolemia (Goldstein and Brown, "Regulation of the mevalonate pathway" *Nature* 343:425-430, 1990. The "statin" class (*e.g.*, compactin and lovastatin) of reductase inhibitors are safe and effective in lowering total cholesterol levels and preventing the progression and reducing the occurrence of coronary disease events (Endo, "The discovery and development of HMG-CoA reductase inhibitors" *J Lipid Res*

33:1569-1582, 1992; Brown, *et al.*, "Regression of coronary artery disease as a result of intensive lipid-lowering therapy in men with high levels of apolipoprotein B" *N Eng J Med* 323:1289-1298, 1990; Nash, "Meeting national cholesterol education goals in clinical practice--a comparison of lovastatin and fluvastatin in primary prevention" *Am J Cardiol* 78: Suppl. 6A:26-31, 1996. Zaragozic acid inhibits squalene synthetase catalyzing the first step of the pathway committed solely to sterol biosynthesis, but appears not to be currently used in the clinical setting. A novel type of zaragozic acid, ZAD3, also inhibits farnesyl protein transferase and, therefore, protein prenylation as well (Tanimoto, *et al.*, "Inhibitory activity to protein prenylation and antifungal activity of zaragozic acid D3, a potent inhibitor of squalene synthase produced by the fungus, *Mollisia* sp sank 10294" *J Antibiot* 51:428-431, 1998). There appears to be no drug in current clinical use that affects solely cholesterol biosynthesis without affecting other farnesyl-PP-derived products.

Applicants have demonstrated that increased levels of an enzyme of the sterol biosynthetic pathway can be assessed indirectly simply by colony color and/or growth on particular media because changes in nonsense suppression occur. Thus, additional mutations and/or drugs that affect the pathway could be identified by a modification of the screening protocol Applicants describe. The effect of prospective new regulators of pathway enzymes on the suppressor ability of a strain with the ALB1 genotype (*mod5-M2 SUP7 can1-100 ade2-1*) could be assessed by adding them to canavanine-containing or adenine-lacking media and monitoring cell growth. If the presence of a substance enhances colony growth on canavanine or deters colony formation on media lacking adenine or causes red colony color to develop, in comparison to the growth/color observed on media without the test substance, then the substance may be increasing the flow of DMAPP through the mevalonate pathway and decreasing tRNA modification. Likewise, using the *mod5-M2, KR6* allele Applicants have previously shown (Zoladek, *et al.* "Mutations altering the mitochondrial-cytoplasmic distribution of Mod5p implicate the actin cytoskeleton and mRNA 3' ends and/or protein synthesis in mitochondrial delivery" *Mol Cell Biol* 15:6884-6894, 1995) that increased cytosolic Mod5p activity results in acquisition of the ability to grow on media lacking lysine. Therefore, compounds that decrease use of DMAPP by the sterol pathway may also be easily assessed. In this way, potential enhancers and inhibitors of the mevalonate pathway could be initially screened for their efficacy quickly and inexpensively.

## II. Assay Applications

### The Mevalonate Pathway

The mevalonate pathway produces isoprenoids that are vital for diverse cellular functions, ranging from cholesterol synthesis to growth control. Several mechanisms for feedback regulation of low-density-lipoprotein receptors and of two enzymes involved in mevalonate biosynthesis ensure the production of sufficient mevalonate for several end-products. Manipulation of this regulatory system could be useful in treating certain forms of cancer as well as heart disease.

In animal cells, a finely tuned metabolic feedback mechanism involving transcriptional and post-transcriptional inputs maintains a pool of mevalonate pathway intermediates essential for cell survival in addition to maintaining cholesterol homeostasis. Cholesterol, the bulk end product of the pathway in sterologenic tissues, exerts transcriptional control on sequential activities in the mevalonate pathway, principally those catalyzed by HMG CoA synthetase, HMG CoA reductase and farnesyl pyrophosphate synthase. HMG CoA reductase is the rate-limiting enzyme of mevalonate biosynthesis as well as the most highly regulated enzyme in this series. The multivalent regulation of HMG CoA reductase integrates activities at the transcriptional, post-transcriptional and post-translational levels. Sterol-mediated regulation of transcription, the dominant regulatory site in sterologenic tissues, is mediated through the sterol regulatory element (SRE), a promoter-enhancer octanucleotide sequence located in the 5' flanking region of the reductase gene.

### Cholesterol

Since cholesterol is present in all tissues of vertebrates and some invertebrates - crustacea (crab, shrimp) and higher mollusks (*e.g.*, squid) - it follows that carnivores ingest appreciable quantities of cholesterol. Not only food but also the bile and, to a small extent, the intestine contribute to the cholesterol available for absorption. Experimental work suggests that the internal cholesterol pool is heterogeneous because biliary cholesterol is more effectively absorbed than dietary cholesterol.

In rats, biliary cholesterol is primarily absorbed in the proximal half of the small bowel, but dietary cholesterol is absorbed in the distal half. Although these observations suggest much better absorption of endogenous than exogenous cholesterol, actually only slightly more endogenous than exogenous cholesterol is absorbed. Several investigators using an isotopic-equilibrium method, concluded that in normal human subjects the maximal capacity for absorbing dietary cholesterol is about 300 to 500 mg/day. Others, using combined chemical and isotope balance methods, have shown that 30 to 40 % of



dietary cholesterol is absorbed in most adults over an intake range of 40 mg/day to more than 2 g/day.

Although some disagree about the percentage of absorption, all investigators agree that only part of the cholesterol of both origins is absorbed. This in itself is a remarkable fact if one considers the different conditions for cholesterol absorption for the two sources of cholesterol. Bile cholesterol, for example, consists of unesterified cholesterol already solubilized in mixed bile salts (BS) phospholipids (PL) micelles. Exogenous cholesterol is not water soluble and is not absorbed as an FA ester. Pancreatic cholesterol esterase, activated by BS, is capable of splitting cholesterol ester into the free sterol and fatty acid (FA), but this is a rather slow process compared to the hydrolysis of triglycerides (TG). Free cholesterol is dissolved in the oil phase and into micelles. When the oil phase disappears, cholesterol precipitates slowing down the rate of absorption.

That cholesterol is only absorbed via micellar solubilization may explain the relatively poor absorption of exogenous cholesterol in healthy subjects, but it does not explain why endogenous cholesterol is poorly absorbed also. It is most likely that cholesterol absorption from micelles slows down considerably when monoglycerides (MG), FA, and PL have disappeared from the micelle because their rate of absorption is much faster and BS micelles alone are poor detergents. Chijiwa and Linscheer's (Chijiwa and Linscheer, "Effect of intraluminal pH on cholesterol and oleic acid absorption from micellar solutions in rat" *Am J Physiol* 246:G492-G499, 1984) observation that absorption rates of cholesterol are much higher at the lower pH of the duodenum and proximal jejunum than at the higher pH in the distal half of the small bowel suggests that this may be a contributing factor.

The physiological malabsorption of cholesterol (related to its very low solubility in water) is essential for the intestinal excretion mechanism of cholesterol. Cholesterol is also the substrate for BS synthesis, but reabsorption of BS is highly efficient and the amount of BS excreted in feces is not the main excretory pathway of cholesterol.

The normal intake of cholesterol (200 to 600 mg/day) accounts for only a small fraction of the total cholesterol transported, and because its contribution to the pool is small, no saturation will be apparent for the absorption of dietary cholesterol. Although individual responses of blood cholesterol show marked variations when animals and humans are exposed to increased amounts of dietary cholesterol and some may even show no clear change, the majority will react with an increase in concentration of blood (and tissue) cholesterol. Apparently no adequate compensatory mechanism exists for the augmented absorption by the reduction of endogenous cholesterol synthesis and for the increased excretion via the bile.

Although an increase dietary cholesterol load may increase serum cholesterol by approximately 15 %, most serum cholesterol is the product of endogenous synthesis. In view of the limited effect of dietary cholesterol on total serum cholesterol, therefore, a method for screening compounds regulating endogenous *de novo* cholesterol synthesis is paramount.

## Cancer

Elucidation of the genes involved in cellular proliferation and the underlying mechanisms that promote and inhibit cell cycle progression has required the use of simple, genetically manipulatable model systems. The present invention makes possible the ability to screen for compounds that may have chemotherapeutic use without the need to run complex genetic experiments. It also allows for the screening of compounds that are agonistic or antagonistic to a pathway known to influence tumor growth and survival.

Mutant *ras* oncogenes and alterations in the mitogenic signaling pathways that they regulate are associated with a wide variety of solid tumors and leukemias for which existing chemotherapeutics have limited utility. Of the possible approaches to inhibit Ras function, much attention has focused on a postranslational modification, farnesylation, which is required for the subcellular localization of Ras to the plasma membrane and is critical to Ras cell-transforming activity. Inhibitors of the enzyme that catalyzes Ras farnesylation, farnesyl-protein transferase (FPTase), have been developed as chemotherapeutics thereby showing the potential use of this approach for the development of anticancer drugs (Gibbs and Oliff, "The potential of farnesyltransferase inhibitors as cancer chemotherapeutics" *Annu. Rev. Pharmacol. Toxicol.* 37:143-166, 1997). However, the identification of these agents by current technologies is a slow and tedious process. The present invention will allow for the high-through put screening of compounds that have potential use as chemotherapeutics.

## Obesity

The present invention is also useful for the screening of compounds that may be beneficial in the control of obesity. Metabolic flux in the mevalonic acid pathway depends on the activity of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase, which regulates the first part of the pathway and the overall synthesis of cholesterol (Goldstein and Brown, "Regulation of the mevalonate pathway" *Nature* 343:425-430, 1990). Experiments with inhibitors of this enzyme have shown that protein prenylation is preserved under conditions that drastically reduce the biosynthesis of cholesterol, suggesting there are regulatory mechanisms acting downstream HMG-C0A

reductase to preserve the supply of FPP and GGPP, the isoprenoids used in protein isoprenylation (Sinensky, *et al.*, "Differential inhibitory effects of lovastatin on protein isoprenylation and sterol synthesis" *J Biol Chem* 265:19937-19941, 1990). GGPP is over expressed in multiple tissues of the *ob/ob* mouse. It is also expressed in increased amounts during adipocyte differentiation in culture. Mammalian GGPP synthase catalyzes the synthesis of FPP and GGPP, the two isoprenoids that take part in protein prenylation. This suggests that this enzyme, by itself or in combination with FPP synthase, serves as a favored point for regulation of prenyl group production for protein prenylation. This protein appears highly regulated during adipogenesis and is overexpressed in an animal model of obesity. In this regard, compounds that regulate the mevalonate pathway may be useful in the regulation of downstream products of the pathway that influence adipogenesis and obesity.

## Plants

The present invention is also useful for the detection and identification of compounds that may be agonistic or antagonistic to the mevalonate pathway in plants. As in yeast, dimethylallyl pyrophosphate (DMAPP) is the substrate of both isopentenyladenasine ( $i^6A$ ) synthesis and a multitude of down stream products from the mevalonate pathway. The mevalonate pathway in plants is responsible for the production of compounds that are necessary for photosynthesis (carotenoids, chlorophylls, phylloquinone, plastoquinone), respiration (ubiquinone, cytochrome a), membrane architecture (sterols and triterpenoids), regulation of growth and development (gibberellic acids, abscisic acid, brassinosteroids, certain cytokinins), defense against pathogen attack (numerous isoprenoid phytoalexins), exchange of signals and prenylation of proteins (allowing for membrane targeting and cell cycle regulation) (Bach, "Some new aspects of isoprenoid biosynthesis in plants - A review" *Lipids* 30:191-202, 1995). Once compounds are found that regulate the mevalonate pathway, they may be further screened by conventional assays known to those in the art for use in various capacities. In this regard, the present invention is useful for the screening of compounds that may be beneficial as regulators of photosynthesis, membrane integrity, plant growth (*e.g.*, fertilizers) and protection (*e.g.*, insecticides, fungicides, antimicrobials and antivirals).

## Fungi

Fungi are single celled living forms of life which inhabit the land, air and waters of the earth. They are more highly developed than bacteria and viruses. It is estimated that there are over 500,000 different species. Fungi have existed on earth hundreds of

millions of years and, quite remarkably, have experienced little genetic change during that period of time. Viable fungi can grow from spores which have been dormant for thousands of years, such as has been observed in spores which were found in Egyptian tombs.

5           Single fungal cells can only be seen under the microscope, but a colony of these cells makes a visible presence in the form of mushrooms, toad stools and molds on food or elsewhere. While plants, animals and humans are alive and well, the fungi around them are usually unable to overcome the natural defense mechanisms which higher forms of life possess. Once death of the living organism has occurred, however, the fungi  
10       become the principle undertakers and managers. They are instrumental in reducing all that has ever lived into the molecules from which they were assembled.

          Unfortunately, though, there is one exception to this simple balanced equation of life and death and that is that the fungi can also attack living cells. At its most simplistic perspective, one has many fungi entering the intestinal tract, the nose and lungs, and  
5       organs exposed to the outside world. Though animals generally do not develop an infection from such intrusions, some persons might contract a fungal infection such as "athlete's foot" or "ring worm" on the skin.

          At the opposite extreme is the patient with AIDS who faces major life-threatening fungal infections because the immune system has lost its ability to protect the body from organisms which invade the body, such as fungi. In between these extremes are fungal  
20       infections associated with diseases such as diabetes, cancer as well as conditions which include intra- and inter-specific cross infections. Two groups of fungi that are of medical importance are yeast and molds. Both are nearly ubiquitous and species of both can cause infections in mammals. Common ailments attributed to fungi are yeast infections and various fungal skin infections. More severe medical conditions can occur if internal  
25       infections arise, e.g., after surgery.

          Additionally, fungi produce mycotoxins. Mycotoxins are biologically active substances that have evolved as a protective mechanism for the fungus. Some mycotoxins have become useful to humans. Antibiotics, such as penicillin, are examples.  
30       However, many mycotoxins can be harmful or deadly. Fungal infections, therefore, are not only limited to the site of infection, but, through the production of mycotoxins, can adversely affect the entire organism. For example, mycotoxins have been linked to cancer. Therefore, even if the fungal infection is cleared, the patient may still suffer long term consequences as a result of the infection.

35       This invention contemplates the screening of compounds that can be useful as anti-fungal agents for both animals and plants. To be useful as an anti-fungal therapeutic,

the desired compound would have to be toxic to the fungus without causing undue negative effects on the patient. The invention, by virtual of the ubiquitousness of the mevalonate pathway, provides a screen where compounds that are potential antifungal therapeutics can be identified. Additionally, the present invention will be useful in the identification of compounds that may be used to eliminate yeast as an infectious agent.

## DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

Generally, the nomenclature used hereafter and the laboratory procedures in cell culture, molecular genetics, and nucleic acid chemistry and hybridization described below are those well known and commonly employed in the art. Standard techniques are used for recombinant nucleic acid methods, polynucleotide synthesis, and microbial culture and transformation (*e.g.*, electroporation, lipofection). Generally enzymatic reactions and purification steps are performed according to the manufacturer's specifications. The techniques and procedures are generally performed according to conventional methods in the art and various general references (see, generally, Sambrook et al. *Molecular Cloning: A Laboratory Manual*, 2d ed. (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., and *Current Protocols in Molecular Biology* (1996) John Wiley and Sons, Inc., N.Y., which are incorporated herein by reference) which are provided throughout this document. All the information contained therein is incorporated herein by reference.

Oligonucleotides can be synthesized on an Applied BioSystems oligonucleotide synthesizer [for details see Sinha *et al.*, *Nucleic Acids Res.* 12:4539 (1984)], according to specifications provided by the manufacturer. Complementary oligonucleotides are annealed by heating them to 90°C in a solution of 10 mM Tris-HCl buffer (pH 8.0) containing NaCl (200 mM) and then allowing them to cool slowly to room temperature. For binding and turnover assays, duplex DNA is purified from native polyacrylamide (15% w/v) gels. The band corresponding to double-stranded DNA is excised and soaked overnight in 0.30 M sodium acetate buffer (pH 5.0) containing EDTA (1 mM). After soaking, the supernatant is extracted with phenol/chloroform (1/1 v/v) and precipitated with ethanol. DNA substrates are radiolabeled on their 5'-OH group by treatment with [ $\gamma$ -<sup>32</sup>P]ATP and T4 polynucleotide kinase. Salts and unincorporated nucleotides are removed by chromatography on Sephadex G columns. Likewise, the cell culture methods described below are also well known in the art.

The present invention relates to the screening of compounds that are agonistic or antagonistic to the mevalonate pathway. Such compounds may be further screened for effectiveness in regulating the production of various end products of the mevalonate

pathway. The regulation of these products may be instrumental in the control or treatment of clinical disease states. For example, such compounds may regulate vascular tone and endothelial function including vascular smooth muscle proliferation, plaque stabilization associated with atherosclerotic lesions and associated cholesterol regulation, prevention of glomerular injury in renal disease, treatment of malignancies and prevention of rejection in organ transplant. The current class of drugs, the statins, used for the control of cholesterol levels exert a physiological effect via regulation of the mevalonate pathway. However, these drugs do not all perform equally in terms of modulating cholesterol levels or in other therapeutic properties. In this regard, new drugs or classes of drugs may be more effective for the various therapeutic purposes listed here or for other therapeutic applications.

The present invention is not limited to any particular mechanism or embodiment. In one embodiment, the  $i^6A$  modification of tRNA catalyzed by Mod5p promotes the efficiency of *SUP7* or *SUP11* tRNAs in the suppression of UAA nonsense mutations by insertion of tyrosine at positions in the growing peptide corresponding to UAA stop codons (Laten *et al.*, "Isopentenyladenosine deficient tRNA from an antisuppressor mutant of *Saccharomyces cerevisiae*" *Nucleic Acids Res.* 5:4329-4342). The extent to which this nonsense suppression occurs during translation in the cytoplasm, can be used as an indication of the amount of Mod5p and isopentenylated tRNA in the cytoplasm.

The *can1-100*, *ade2-1*, and *lys2-1* alleles contain UAA nonsense mutations. *CAN1* encodes an arginine permease that allows the uptake of the arginine analog canavanine (Whelan *et al.*, "The *CAN1* locus of *Saccharomyces cerevisiae*: fine-structure analysis and forward mutation rates" *Genetics* 91:35-51, 1979). Canavanine interferes with the process of translation and cells can not grow in its presence. Therefore, cells with wild-type *CAN1* are sensitive to canavanine ( $Can^s$ ), but cells with the mutant *can1-100*, that does not encode a properly functioning permease, are resistant to canavanine ( $Can^r$ ) and can grow in its presence. Cells with  $i^6A$  completely modified *SUP7* or *SUP11* tRNAs can not grow in the presence of canavanine, whereas cells lacking  $i^6A$  on these tRNAs are able to grow in the presence of this drug. Cells with intermediate levels of  $i^6A$  show intermediate levels of growth in the presence of the drug.

*ADE2* encodes an enzyme involved in the synthesis of adenine. Cells with *ade2-1* turn red in color, due to accumulation of a metabolic intermediate, and fail to grow on defined media lacking adenine ( $Ade^-$ ), while cells producing a functional Ade2p can grow on such media and are white ( $Ade^+$ ) (Rasse-Messenguy and Fink, "Temperature-sensitive nonsense suppressors in yeast" *Genetics* 75:459-464, 1973). Cells with the *ade2-1* allele and sufficient  $i^6A$  modified suppressor tRNA can grow in the absence of

exogenous adenine and generate white colonies on rich media whereas cells with insufficient i<sup>6</sup>A modified suppressor tRNA are unable to grow in the absence of exogenous adenine and generate red colonies on rich media. Cells with intermediate levels of i<sup>6</sup>A modified tRNA have intermediate phenotypes in colony color and intermediate rates of growth in the absence of exogenous adenine.

*LYS2* encodes an enzyme involved in the biosynthesis of lysine. Mutations in this gene prevent growth on media lacking exogenous lysine. The *lys2-1* nonsense allele can be suppressed by *SUP7* or *SUP11* provided that the encoded nonsense codon suppressing tRNAs possess an i<sup>6</sup>A modification at position 37. Cells with no Mod5p lack i<sup>6</sup>A modified tRNA and are unable to grow in the absence of exogenous lysine whereas cells with active cytosolic pools of Mod5p can grow in the absence of this amino acid. Cells with intermediate levels of i<sup>6</sup>A modified tRNA have intermediate abilities for growth on media lacking lysine.

When a cell contains the wild-type *MOD5*, encoding both isoforms of Mod5p, the *SUP7* or *SUP11* pools are fully modified. Such strains efficiently suppress the *can1-100*, *ade2-1* and *lys2-1* and they are Can<sup>s</sup>, Ade<sup>+</sup>, and Lys<sup>+</sup>. Cells which have the mutant *mod5-1* allele contain approximately 1.5% of the amount of isopentenyl adenosine present in *MOD5* cells, so little suppression of *can1-100*, *ade2-1* and *lys2-1* will occur, making them Can<sup>r</sup>, Ade<sup>-</sup> and Lys<sup>-</sup>. Cells with the *mod5-M2* allele (strain ALB1, Figure 2), which contains a mutation at codon 12 preventing the initiation of Mod5p-II translation and allowing only Mod5p-I production, have about 50% of the *SUP7* pool modified (Figure 3) resulting in intermediate levels of suppression. Such cells show intermediate growth on canavanine (Can<sup>r/s</sup>) and on media lacking adenine (Ade<sup>+/-</sup>) and generate pink colonies on rich media. Cells with the *mod5-M2KR6* allele (Boguta *et al.*, "Subcellular locations of MOD5 proteins: mapping of sequences sufficient for targeting to mitochondria and demonstration that mitochondrial and nuclear isoforms commingle in the cytosol" *Mol Cell Biol.* 14:2298-2306, 1994; Zoladek *et al.*, "Mutations altering the mitochondrial-cytoplasmic distribution of Mod5p implicate the actin cytoskeleton and mRNA 3' ends and/or protein synthesis in mitochondrial delivery" *Mol Cell Biol* 15:6884-6894, 1995) have a very small cytosolic pool of i<sup>6</sup>A-tRNA and they are unable to grow in the absence of lysine (Figure 2).

In the process of searching for genes that caused decreased cytosolic pools of Mod5p-I, *ERG20* was found (Benko *et al.*, "Competition between a sterol biosynthetic enzyme and tRNA modification in addition to changes in the protein synthesis machinery causes altered nonsense suppression" *PNAS* 97:61-66, 2000). *ERG20* encodes farnesyl diphosphate synthetase. This enzyme catalyzes the formation of geranyl pyrophosphate

and farnesyl pyrophosphate by the condensation of units of dimethylallyl pyrophosphate (DMAPP) and its isomer isopentenyl pyrophosphate (IPP). DMAPP is the same molecule used by Mod5p as a substrate in the synthesis of i<sup>6</sup>A in tRNA (Brown and Goldstein, "Multivalent feedback regulation of HMG CoA reductase, a control mechanism coordinating isoprenoid synthesis and cell growth" *J Lipid Res.* 21:505-517, 1980. The applicants proposed that Erg20p over-expression competed for the DMAPP normally channeled to tRNA modification by Mod5p and showed that when Erg20p is highly expressed the level of i<sup>6</sup>A modification is reduced more than 3-fold. This results show that the mevalonate pathway and the tRNA pathway are in competition and small differences in i<sup>6</sup>A modification of tRNA that result from competition and flux in the sterol biosynthesis pathway can be detected by simple growth/color assays that score nonsense suppression.

Assays for detecting the ability of agents to inhibit or enhance flux through the mevalonate pathway provide for facile high-throughput screening of agent banks (e.g., compound libraries, peptide libraries, and the like) to identify antagonists or agonists. Such mevalonate pathway antagonists and agonists may be further developed as potential therapeutics and diagnostic or prognostic tools for diverse types of clinical diseases.

#### **1. Screens to identify Agonists or Antagonists of Mevalonate Pathway Activity**

There are several different approaches contemplated by the present invention to look for molecules that specifically are agonistic or antagonistic to the mevalonate pathway.

##### **i. Colorimetric Assays**

In one embodiment, yeast cells of the stains ALB1 and ALB8 are cultured, exposed to one or more compounds and color is determined visually. In another embodiment, cells are lysed and color is determined with a spectrophotometer.

##### **ii. Growth Assays**

In one embodiment, yeast cells of the stains ALB1, ALB8, MD14A, T8-ID are cultured, exposed to one or more compounds and growth is determined visually. In another embodiment, cells are suspended in an isotonic solution and cell growth is determined with a spectrophotometer or with a flow cytometer.

##### **iii. Immunoprecipitation and Western blot**

After the generation of antibodies to i<sup>6</sup>A, cells are lysed and then incubated with the antibodies. Antibodies interact with the i<sup>6</sup>A and any associated proteins can then be pulled down with protein-A Sepharose or protein-G Sepharose beads or detected by Western blot, using standard techniques.



## EXPERIMENTAL

The following examples are intended to illustrate, but not limit, the present invention. Additionally, the following examples employ standard molecular biological, microbiological and cell culture procedures. These techniques and procedures are generally performed according to conventional methods in the art (see, generally, Sambrook et al. *Molecular Cloning: A Laboratory Manual*, 2nd ed. (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., and *Current Protocols in Molecular Biology* (1996) John Wiley and Sons, Inc., N.Y., which are incorporated herein by reference).

### EXAMPLE I

**Identification of Plasmids Causing Reduced Nonsense Suppression and Sequencing of Inserts of Library Clones.** The applicants employed the *can1-100* and *ade2-1* alleles in a *mod5-M2* background (strain ALB1) to identify plasmids causing alterations in nonsense suppression. The strains were made as follows: Strain ALB1 (MAT $\alpha$  *mod5-M2 SUP7 ade2-1 can1-100 leu2-3, -112 lys1-1 lys2-1 trp1 ura3-1*) was created from MD14A (Gillman, *et al.*, "MOD5 translation initiation sites determine N6-isopentenyladenosine modification of mitochondrial and cytoplasmic tRNA" *Mol Cell Biol* 14:2298-2306, 1994) by the replacement of the genomic *mod5-1* through a pop-in/pop-out procedure (Rothstein, "Targeting, disruption, replacement, and allele rescue: integrative DNA transformation in yeast" *Methods Enzymol* 194:281-301, 1991). Plasmid YCfmod5-M2 (Gillman, *et al.*, "MOD5 translation initiation sites determine N6-isopentenyladenosine modification of mitochondrial and cytoplasmic tRNA" *Mol Cell Biol* 14:2298-2306, 1994) was used as the donor of the M2 mutation. The *ARS1* and *CEN4* sequences were removed from YCfmod5-M2 by a *Bgl*III/*Sma*I digestion. The *Bgl*III end of the remaining plasmid was filled in using Klenow DNA Polymerase, and then ligated to the *Sma*I blunt end. The resulting plasmid was linearized by *Sna*BI just upstream of the *mod5-M2* ORF and then transformed into MD14A. Integration of the plasmid was selected for by growth on media lacking uracil. Resulting cells contained two *mod5* alleles: *mod5-1* and *mod5-M2*. Cells in which intrachromosomal homologous recombination and loss of the *mod5-1* mutation occurred were selected for by growth on media containing 5-fluoro-orotic acid (5-FOA), due to concomitant loss of *URA3*, and for the ability to grow on media lacking adenine. The presence of the remaining *mod5-M2* allele was confirmed by DNA blot hybridization.

Strain ALB8 (*MATa SUP7 can1-100 ade2-1 leu2-3, -112 lys1-1 lys2-1 trp1 mod5::TRP1 ura3-1::MOD5*) was created through integration of a plasmid containing *MOD5* at the *ura3-1* locus of strain MT-8 (Gillman, *et al.*, "MOD5 translation initiation sites determine N6-isopentenyladenosine modification of mitochondrial and cytoplasmic tRNA" *Mol Cell Biol* 11: 2382-2390, 1991). Plasmid YCfMOD5 (Gillman, *et al.*, "MOD5 translation initiation sites determine N6-isopentenyladenosine modification of mitochondrial and cytoplasmic tRNA" *Mol Cell Biol* 11: 2382-2390, 1991) was used as the donor of *MOD5*. The *ARS1* and *CEN4* sequences were removed as described above, and the resulting plasmid was linearized with *NcoI* to target it to *ura3-1*. MT-8 cells that acquired the plasmid were selected on media lacking uracil. Correct integrants were assessed by DNA blot hybridization. Strain SL680-9C (*MATa asu9-1sup45-2 ura3-52 can1-132 leu2-1 lys2-1 met8-1 trp1-1* or *trp1-289*) (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994) was obtained from S. Liebman.

Yeast strains were propagated in YEPD media or synthetic complete (SC) media lacking certain nutrients to select for the presence of plasmid and to assay suppression. Canavanine sulfate salt (SIGMA) was added to media lacking arginine to a final concentration of 30 µg/ml. Yeast strains were transformed following a modified TRAF0 protocol (Gietz, *et al.*, "Improved method for high efficiency transformation of intact yeast cells" *Nucleic Acids Res* 20:1425, 1992). Additionally, the following library DNAs and plasmids were used: DNA library based in YEp13 (Nasmyth and Tatchell, "The structure of transposable yeast mating type loci" *Cell* 19:753-764, 1980), a genomic DNA library based in YEp24 (Carlson and Botstein, *et al.*, "Two differentially regulated mRNAs with different 5' ends encode secreted with intracellular forms of yeast invertase" *Cell* 28:145-154, 1982), a cDNA library based in pMac561, a 2µm vector with the *ADHI* promoter (McKnight and McConaughy, "Selection of functional cDNAs by complementation in yeast" *Proc Natl Acad Sci USA* 80:4412-4416) and a cDNA library based in pRS316 (Liu, *et al.*, "Construction of a GAL1-regulated yeast cDNA expression library and its application to the identification of genes whose overexpression causes lethality in yeast" *Genetics* 132:665-673, 1992).

YEpMOD5 contains the 1.8 kb DNA fragment encoding wild-type *MOD5* gene and regulatory regions in the YEp24 vector (Dihanich, *et al.*, "Isolation and characterization of *MOD5*, a gene required for isopentenylation of cytoplasmic and mitochondrial tRNAs of *Saccharomyces cerevisiae*" *Mol Cell Biol* 7:177-184, 1987). pRH127-3, a gift from R. Wright, is a 2micron-based plasmid that contains a truncated

*HMG1* gene, that causes increased HMG-CoA reductase activity and increased levels (~10X greater) of intermediates in the mevalonate pathway (Donald, *et al.*, "Effects of overproduction of the catalytic domain of 3-hydroxy-3-methylglutaryl coenzyme A reductase on squalene synthesis in *Saccharomyces cerevisiae*" *Appl Environ Microbiol* 63:3341-3344, 1997).

Library plasmids were isolated from yeast by the method of Ward (Ward, "Single-step purification of shuttle vectors favor yeast for high frequency back-transformation into *E. coli*" *Nucleic Acids Res* 18:5319, 1990). DNAs were sequenced by either the chain termination method (Sanger, *et al.*, "DNA sequencing with chain-terminating inhibitors" *Proc Natl Acad Sci USA* 74:5463-5467, 1977) with Sequenase Version 2.0 DNA Sequencing Kit (United States Biochemical) or by automated cycle sequencing performed in the Pennsylvania State University College of Medicine Macromolecular Core Facility. Nucleotide sequences were identified by a BLAST (Altschul, *et al.*, "Basic local alignment search tool" *J Mol Biol* 215:403-410, 1990) search at the *Saccharomyces* Genome Database BLAST server (<http://genome-www2.stanford.edu/cgi-bin/SGD/nph-blast2sgd/>).

Mod5-M2 is an allele of *MOD5* that encodes normal levels of mitochondrial but reduced levels of cytosolic Mod5p activity. In strains, with this allele cytosolic tRNAs are only partially modified with i<sup>6</sup>A (60%; Gillman, *et al.*, "*MOD5* translation initiation sites determine N6-isopentenyladenosine modification of mitochondrial and cytoplasmic tRNA" *Mol Cell Biol* 11:2382-2390, 1991) and further alterations in Mod5p levels are readily assessed by changes in nonsense suppression (Zoladek, *et al.*, "Mutations altering the mitochondrial-cytoplasmic distribution of Mod5p implicate the actin cytoskeleton and mRNA 3' ends and/or protein synthesis in mitochondrial delivery" *Mol Cell Biol* 15:6884-6894, 1995). The *can1-100* and *ade2-1* alleles contain UAA nonsense mutations. *CAN1* encodes an arginine permease that allows the uptake of the arginine analog canavanine (Whelan, *et al.*, "The *CAN1* locus of *Saccharomyces cerevisiae*: fine-structure analysis and forward mutation rates" *Genetics* 91:35-51, 1979), and cells cannot grow in its presence. Therefore, cells with wild-type *CAN1* are sensitive to canavanine (Can<sup>s</sup>), but cells with the mutant *can1-100* are resistant to canavanine (Can<sup>r</sup>). Cells containing *SUP7* tRNAs completely modified with i<sup>6</sup>A have functional arginine permease and, therefore, cannot grow in the presence of canavanine, whereas cells lacking i<sup>6</sup>A on their tRNAs are able to grow in the presence of this drug. Cells with intermediate levels of i<sup>6</sup>A show intermediate levels of growth in the presence of the drug.

*ADE2* encodes an enzyme involved in the synthesis of adenine. Cells with *ade2-1* turn red in color and fail to grow on defined medium lacking adenine (Ade<sup>-</sup>), whereas

cells producing functional Ade2p can grow on such medium (Ade<sup>+</sup>) and are white (Rasse-Messenguy and Fink, "Temperature-sensitive nonsense suppressors in yeast" *Genetics* 75:459-464, 1973). Cells with the *ade2-1* allele and sufficient i<sup>6</sup>A-modified suppressor tRNA can grow in the absence of exogenous adenine and generate white colonies on rich medium, whereas cells with insufficient i<sup>6</sup>A-modified tRNA are unable to grow in the absence of exogenous adenine and generate red colonies on rich medium. Cells with intermediate levels of i<sup>6</sup>A modified tRNA have intermediate phenotypes in colony color and intermediate rates of growth in the absence of exogenous adenine.

Transformants able to grow on canavanine were selected from approximately 1.5 million independent ALB1 (relevant genotype: *SUP7 ade2-1 can1-100 mod5-M2*) transformed cells. One hundred and fifty-eight candidates that exhibited growth on medium containing canavanine were tested for lack of growth on medium lacking adenine to eliminate those gaining canavanine resistance because of additional nonsuppressible mutations in *can1-100*.

For each of the 71 canavanine-resistant candidates that also were unable to grow well on medium lacking adenine or were red-pink in color, an assay was performed to determine whether these phenotypes required the presence of the library plasmid. Thirty-three candidates showed dependence of suppression levels upon the presence of a plasmid. Analysis of the plasmids in these cells resulted in the identification of 11 with unique restriction patterns. These plasmids were retransformed into ALB1 and the transformants were reassayed for canavanine resistance and adenine prototrophy. One plasmid did not confer either of these phenotypes. Four pRS316 library clones were still able to prevent growth on medium lacking adenine, and six of the remaining clones still could produce both desired phenotypes in ALB1 as confirmed by a second set of co-loss experiments (Table 1). Sequencing of the inserts of these latter 10 clones led to the identification of five different genes: *SAL6*, *ARC1*, *TEF4*, *YDL219W*, and *ERG20*.

*ERG20*, which encodes farnesyl diphosphate synthetase, was found once in each of the YEp13, pMac561, and pRS316 libraries. *SAL6* cDNA, encoding a protein phosphatase, was discovered four times in the pRS316 library. *ARC1*, *TEF4*, and the putative ORF *YDL219w* were single candidates from the pRS316 library (Table 1). The pRS316 library is constructed such that the cloned genes are regulated by the *GAL1* promoter. Thus, transcription is induced by galactose and repressed by glucose in the media. The effects of the identified genes upon nonsense suppression were dependent on having galactose in the medium (Table 1), showing that the phenotypes are a result of their expression. These clones were used for the examples that follow.

**TABLE 1. Growth of Strains Containing Plasmids**

Strain	Growth on canavanine	Growth/colony color on SC-adenine	
	Galactose	Glucose	Galactose
MD14A	++	- r	- r
pRSvector/ALB1	-	+ w	+ w
pRSSAL6/ALB1	+	+ w	+/- p
pRSESG20/ALB1	+	+ w	+/- p
pRSARC1/ALB1	+/-	+ w	+/- p
pRSYDL219w/ALB1	+/-	+ w	+/- p
pRSTEF4/ALB1	+/-	+ w	+/- p
++, Very strong growth; +, strong growth; +/-, poor growth; -, no growth; r, red; p, pink; w, white.			

## EXAMPLE 2

### **SAL6, TEF4, and YDL219w Confer Antisuppressor Phenotypes.**

*SUP45* encodes a translation termination factor, and mutation of this gene can result in omnipotent suppression (Stansfield and Tuite, "Polypeptide chain termination in *Saccharomyces cerevisiae*" *Curr Genet* 25:385-395, 1994; Stansfield, *et al.*, "The products of the SUP45 (eRF1) and SUP35 genes interact to mediate translation termination in *Saccharomyces cerevisiae*" *EMBO J* 14:4365-4373, 1995). When *SAL6* is overexpressed in a yeast strain containing *sup45-2*, an anti-suppressor phenotype occurs (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994). This suggested that the canavanine resistance conferred upon ALB1 by high levels of Sal6p also may be due to antisuppression. Therefore, all of the candidate genes were tested for effects upon translation by assessing whether they scored as antisuppressors of *sup45-2*.

SL680-9C (relevant genotype *sup45-2 asu9-1 leu2-1 met8-1 ura3-52*) is capable of growing on medium lacking methionine because the mutant *sup45* permits translation through nonsense mutations such as *met8-1* (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994; Stansfield

d and Tuite, "Polypeptide chain termination in *Saccharomyces cerevisiae*" *Curr Genet* 25:385-395, 1994; Stansfield, *et al.*, "The products of the SUP45 (eRF1) and SUP35 genes interact to mediate translation termination in *Saccharomyces cerevisiae*" *EMBO J* 14:4365-4373, 1995). When *SAL6* is overexpressed in a yeast strain containing *sup45-2*, an anti-suppressor phenotype occurs (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994). The *asu9* antisuppressor allele serves to weaken the effects of *sup45* and permits better growth of cells with both mutations (Vincent, *et al.*, "The yeast translational allosuppressor, *SAL6*: a new member of the PP1-like phosphatase family with a long serine-rich N-terminal extension" *Genetics* 138:597-607, 1994; Stansfield and Tuite, "Polypeptide chain termination in *Saccharomyces cerevisiae*" *Curr Genet* 25:385-395, 1994).

**TABLE 2. Growth of SL680-9C cells overexpressing candidate genes**

Candidate gene/ORF	Growth on SC-uracil-methionine (galactose)	Growth/colony color on SC-adenine (glucose)
None (vector only)	+	+
<i>SAL6</i>	-	+
<i>ARC1</i>	+	+
<i>TEF4</i>	+/-*	+
<i>ERG20</i>	+	+
<i>YDL219w</i>	+/-	+
Cells were grown at 28 °C for 5-8 days; results are for three experiments. +, strong growth; +/-, poor growth; -, no growth. *Galactose vs. glucose growth difference sometimes not apparent.		

SL680-9C cells transformed by each of the candidate genes in the pRS316 vector were analyzed for growth on galactose-containing or glucose-containing medium lacking uracil and methionine. The data (Table 2) indicate that overexpressed *SAL6* and, to a lesser degree, *TEF4* and *YDL219w* cause antisuppression, and, therefore, loss of suppression of the *can1-100* and *ade2-1* in *ALB1* is likely Mod5p-I-independent. This

phenotype was not observed consistently in the cells containing large amounts of Tef4p. Neither Arc1p nor Erg20p in high levels affected the suppression of *met8-1* in SL680-9C, and, therefore, each probably affects *can1-100* and *ade2-1* suppression in ALB1 via another route, likely by alteration of Mod5p-I or tRNA activities or location.

### EXAMPLE 3

**Overexpression of ERG20 Results in the Reduction of Isopentenyl Adenosine Present in tRNA.** The protein encoded by *ERG20*, farnesyl diphosphate synthetase, catalyzes the formation of geranyl pyrophosphate and farnesyl pyrophosphate by the condensation of units of DMAPP and its isomer isopentenyl pyrophosphate (IPP). DMAPP is the same molecule used by Mod5p as a substrate in the synthesis of  $i^6A$  in tRNA (Figure 1; Brown and Goldstein, "Multivalent feedback regulation of HMG CoA reductase, a control mechanism coordinating isoprenoid synthesis and cell growth" *J Lipid Res* 21:505-517, 1980; Voet and Voet, *Biochemistry* (Wiley, NY) 1990). It was possible that the large pool of Erg20p produced from the library clones used the DMAPP normally utilized for tRNA modification. This would cause reduced  $i^6A$ -modified tRNA and, hence, reduced function of these tRNAs in nonsense suppression. It was also possible that others of the candidate genes, when overexpressed, inhibited the formation of  $i^6A$  tRNA.

To determine the effect of overexpression of the candidate genes upon  $i^6A$  modification of tRNA, Applicants employed the Mid-western procedure (Rasmussen and Culbertson, "Analysis of yeast trimethylguanosine-capped RNAs by midwestern blotting" *Gene* 182:89-96, 1996) by using anti- $i^6A$  antibody.

Low-molecular-weight RNA was prepared from cells with each plasmid and from control strains of ALB1 with vector only, ALB8, and MD14A. The RNAs were resolved on urea-polyacrylamide gels, transferred to membranes, and probed with anti- $i^6A$  antibody (Figure 2A, upper). Hybridization with a radiolabeled oligonucleotide complementary to part of mature tRNA Tyr was used to assess the relative amount of tRNA in each lane (Figure 2A, lower). Through densitometric analysis, the amount of  $i^6A$  formed in tRNA of each strain as a fraction of the amount formed in the vector-only control strain was determined (Figure 2B).

As expected (Martin and Hopper, "Isopentenylolation of both cytoplasmic and mitochondrial tRNA is affected by a single nuclear mutation" *J Biol Chem* 257:10562-10565, 1982), MD14A cells with the *mod5-1* allele have barely detectable levels of modified tRNA. Also as anticipated, ALB8 cells with a wild-type *MOD5* that encodes both Mod5p-I and Mod5p-II contained higher levels (~35% more) of  $i^6A$ -modified tRNA

than ALB1 cells expressing only Mod5p-I (Figure 2; Gillman, *et al.* "MOD5 translation initiates sites determine N<sup>6</sup>-isopentenyladenasine modification of mitochondrial and cytoplasmic tRNA" *Mol Cell Biol* 11:2382-2390, 1991). The level of i<sup>6</sup>A tRNA in ALB1 overexpressing *SAL6*, *ARC1*, *YDL219w*, or *TEF4* is similar to the level of i<sup>6</sup>A from ALB1 cells with vector only. However, the amount of i<sup>6</sup>A tRNA in ALB1 with excess Erg20p is significantly reduced to approximately 30% of that in ALB1 with vector alone. Overproduction of Erg20p results in reduced levels of i<sup>6</sup>A tRNA and causes a decrease in nonsense suppression.

**TABLE 3. Growth of Strains Containing Multicopy Plasmids**

ALB1 with plasmids	Growth on canavanine	Growth/colony color on SC-adenine
YEp13 + YEpMOD5	-	+ w
YEpERG20 + YEp24	+/-	+/- p
YEpERG20 + YEpMOD5	-	+ w
YEp13 + YEp24	-	+ w
YEp13 + pRH127-3	-	+ w
YEpERG20 + YEp24	+/-	+/- p
YEpERG20 + pRH127-3	+/-	+ w
+,strong growth; +/-, poor growth; -, no growth; p, pink; w, white.		

If overproduction of Erg20p acts via competition with Mod5p-I for a common substrate (Figure 3), then providing additional Mod5p should reverse the loss of suppression. To test this, Applicants transformed ALB1 cells containing YEpERG20, ERG20 in a multi copy vector, with YEpMOD5 and assessed suppression of *ade2-1* and *can1-100*. The results (Table 3) show that suppression indeed is restored by additional Mod5p. Likewise, if Erg20p and Mod5p are in competition for the same substrate, then alterations that increase the substrate should dampen the competition. To test this, Applicants introduced a multicopy vector harboring a mutant *HMG1* gene, pRH127-3,



that causes elevated levels of intermediates of the mevalonate pathway (Donald, *et al.*, "Effects of overproduction of the catalytic domain of 3-hydroxy-3-methylglutaryl coenzyme A reductase on squalene synthesis in *Saccharomyces cerevisiae*" *Appl Environ Microbiol* 63:3341-3344, 1997) into ALB1 containing YEpERG20. Applicants found that increased levels of the intermediates also restored nonsense suppression (Table 3). The data support the model that the mevalonate pathway and the i<sup>6</sup>A tRNA modification pathways are in competition.

Determination of the levels of i<sup>6</sup>A in tRNA. RNA was obtained (Hopper, *et al.*, "Processing of intervening sequences: a new yeast mutant which fails to excise intervening sequences from precursor tRNAs" *Cell* 19:741-751, 1980) from overnight log phase cultures of the following strains: ALB8, MD14A and ALB1 with the pRS316 library plasmids containing *SAL6*, *ARC1*, *TEF4*, *ERG20*, YDL219w or no cDNA insert. The RNA concentration was calculated from optical density measurement.

RNAs were resolved on a 12% polyacrylamide / 7M urea gel. The RNA was transferred to Hybond N+ Nylon Membrane (Amersham) in a Hoefer TE Series Transphor Apparatus containing 1X TAE. The RNA was fixed onto the membrane by UV crosslinking. The membrane was subjected to immunoblotting following the protocol of the Renaissance Western Blot Chemiluminescence Kit (NEN) with slight variation. The primary antibody, anti-i<sup>6</sup>A isolated from rabbit prepared by the method of Senapathy and Jacob (Senapathy and Jacob, "Identification and purification of tRNAs containing N6-(delta 2-isopentenyl) adenosine using antibodies specific for N6-(delta-isopentenyl) adenosine" *J Biol Chem* 256:11580-11584, 1981), was diluted either 1:33.3 or 1:72.5 prior to use. Horseradish peroxidase conjugated anti-rabbit IgG from donkey (Amersham) diluted 1:5000 or 1:2500 served as secondary antibody. The membrane was also utilized for RNA detection using radiolabeled oligonucleotide complementary to a region of mature tRNA<sup>Tyr</sup>.

The intensities of the fluorescent and radioactive signals for each specimen on the immunoblots and RNA blots, respectively were determined by densitometry using a Molecular Dynamics laser scanner with the Discovery Series Quantity One computer software (Protein Database, Inc.). For each sample, the ratio of its signal with respect to that of the signal from the ALB1 with vector only sample was calculated from the immunoblots (Ratio 1) and from the RNA blots (Ratio 2). The average values for Ratio 1 to Ratio 2 for samples from two membranes were calculated. The data show that increased levels of enzyme activity of the sterol biosynthetic pathway can be assessed indirectly simply by colony color and / or growth on particular medium because changes

in nonsense expression occur. Thus, additional mutations and / or drugs that affect the pathway could be identified by the screening protocol described.

#### EXAMPLE 4

5           **Automated assays.** In this experiment yeast cells are cultured as described above in microtiter plates and contact with compounds in order to assess the ability of the compound to modulate the mevalonate pathway. After culture, the cells are lysed *in situ* and the microtiter plates are analyzed with a manual or automated spectrophotographic plate reader. Attachment of the reader to a computer will allow for the near instantaneous  
10 recording and analysis of the data.

          Another experiment will utilize automated flow cytometry. In this assay whole cells are analyzed with an automatic sampling flow cytometer calibrated to detect color and / or emitted fluorescence. Additionally, the relative number of cells between samples will be determined to measure cell growth. In any of these examples robotics will be  
15 used to analyze a large number of samples quickly.

          Downstream products of the mevalonate pathway may be both characterized and quantitated. In addition to the analyses above, effect of the compounds on the mevalonate pathway may be confirmed by analysis of the increased or decreased production of downstream mevalonate pathway products. Such analyses are well known  
20 in the art and include radioimmunoassays (RIA) and high-pressure liquid chromatography (HPLC). In this way, the effected downstream pathway or pathways will be determined.

          It should be evident from the foregoing that the present invention provides novel materials and methods that will permit the rapid screening of compounds for their  
25 agonistic or antagonistic effect on the mevalonate pathway. In addition, the present invention provides novel materials and methods for the determination of existing or promising therapeutic compounds that may have agonistic or antagonistic side effects on the mevalonate pathway.

## CLAIMS

1. A method, comprising:

- 5 a) providing: i) a test compound, ii) a growth media formulated to allow scoring of nonsense suppression in yeast, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p or its homolog as compared to said wild type yeast cells, and wherein said modified yeast cells comprise a gene with a nonsense mutation and a suppressor tRNA gene coding for a tRNA modified with isopentenyl adenosine by Mod5 or its homolog;
- 10 b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and
- 15 c) measuring for growth of said treated portion.

2. The method of Claim 1, wherein said measuring of step (c) comprises examining the color of said yeast cells of said treated portion.

3. The method of Claim 1, wherein said measuring of step (c) comprises comparing said treated portion with said untreated portion, wherein said untreated portion is exposed to said growth media in the absence of said test compound.

4. A method, comprising:

- 25 a) providing: i) a test compound, ii) a growth media lacking adenine, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise an *ADE* gene having a nonsense mutation and a gene coding for a nonsense suppressor tRNA;
- 30 b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and
- c) measuring for growth of said treated portion.

5. The method of Claim 4, wherein said measuring of step (c) comprises examining the color of said yeast cells of said treated portion.

6. The method of Claim 4, wherein said measuring of step (c) comprises comparing said treated portion with said untreated portion, wherein said untreated portion is exposed to said growth media in the absence of said test compound.

7. A method, comprising:

- a) providing: i) a test compound, ii) a growth media lacking adenine, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise an *ADE* gene having a nonsense mutation and a *SUP7* gene coding for a tRNA;
- b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and
- c) measuring for growth of said treated portion.

8. The method of Claim 7, wherein said measuring of step (c) comprises examining the color of said yeast cells of said treated portion.

9. The method of Claim 7, wherein said measuring of step (c) comprises comparing said treated portion with said untreated portion, wherein said untreated portion is exposed to said growth media in the absence of said test compound.

10. A method, comprising:

- a) providing: i) a test compound, ii) a growth media lacking arginine and comprising a canavanine salt, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise a *CAN1* gene having a nonsense mutation and a gene coding for a nonsense suppressor tRNA;
- b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and
- c) measuring for growth of said treated portion.

11. The method of Claim 10, wherein said measuring of step (c) comprises comparing said treated portion with said untreated portion, wherein said untreated portion is exposed to said growth media in the absence of said test compound.

12. A method, comprising:

- a) providing: i) a test compound, ii) a growth media lacking arginine and comprising a canavanine salt, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise a *CAN1* gene having a nonsense mutation and a *SUP7* gene coding for a tRNA;
- b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and
- c) measuring for growth of said treated portion.

13. The method of Claim 12, wherein said measuring of step (c) comprises comparing said treated portion with said untreated portion, wherein said untreated portion is exposed to said growth media in the absence of said test compound.

14. A method, comprising:

- a) providing: i) a test compound, ii) a growth media lacking lysine, and iii) modified yeast cells derived from wild type yeast cells, wherein said modified yeast cells express reduced cytosolic levels of Mod5p as compared to said wild type yeast cells, and wherein said modified yeast cells comprise a *LYS2* gene having a nonsense mutation and a gene coding for a nonsense suppressor tRNA;
- b) exposing a portion of said modified yeast cells to said test compound and said growth media to create a treated portion and an untreated portion; and
- c) measuring for growth of said treated portion.

15. The method of Claim 14, wherein said measuring of step (c) comprises comparing said treated portion with said untreated portion, wherein said untreated portion is exposed to said growth media in the absence of said test compound.

16. The method of Claim 14, wherein said gene coding for said nonsense suppressor tRNA is selected from the group consisting of *SUP7* and *SUP11*.

## ABSTRACT

5

A novel assay for the detection of substances agonistic or antagonistic to the mevalonate pathway are disclosed.

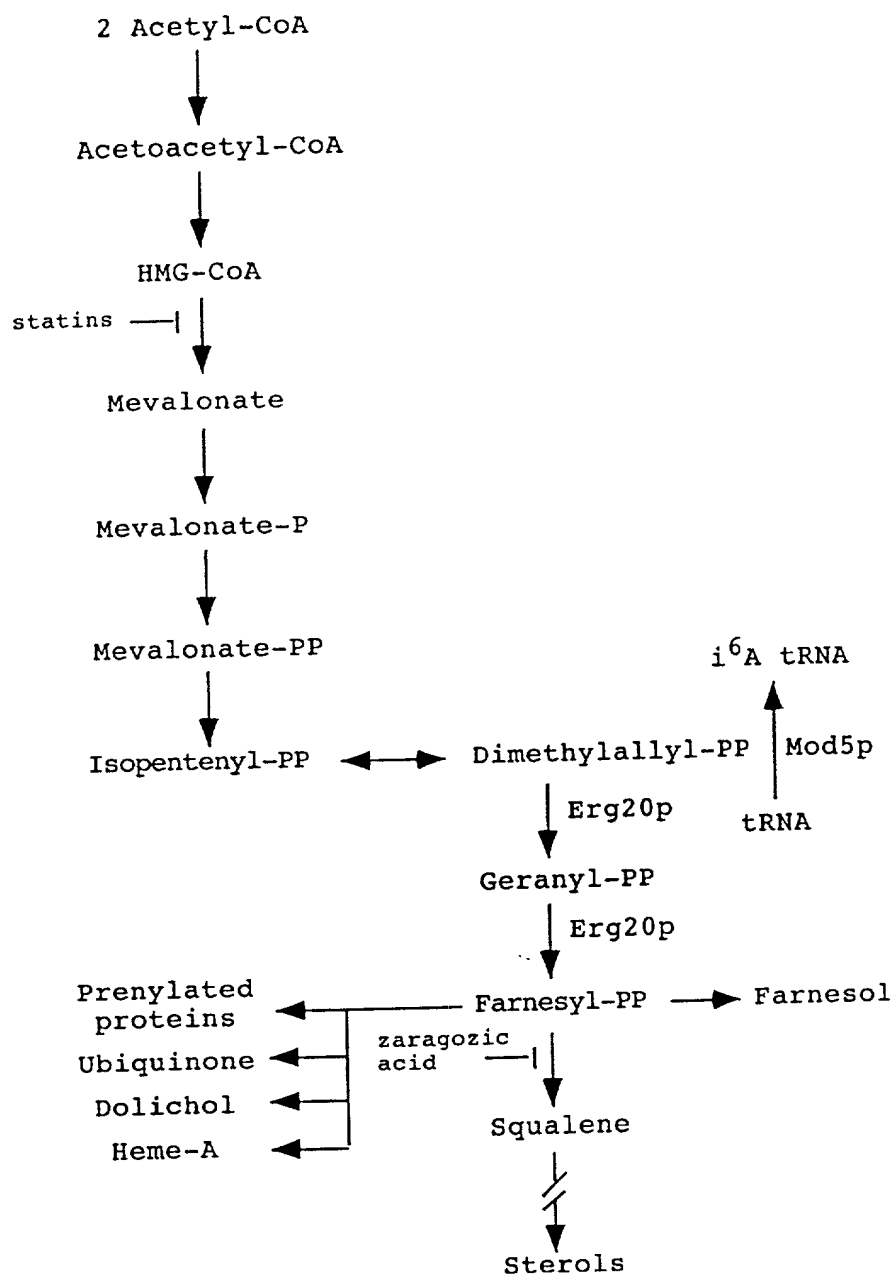
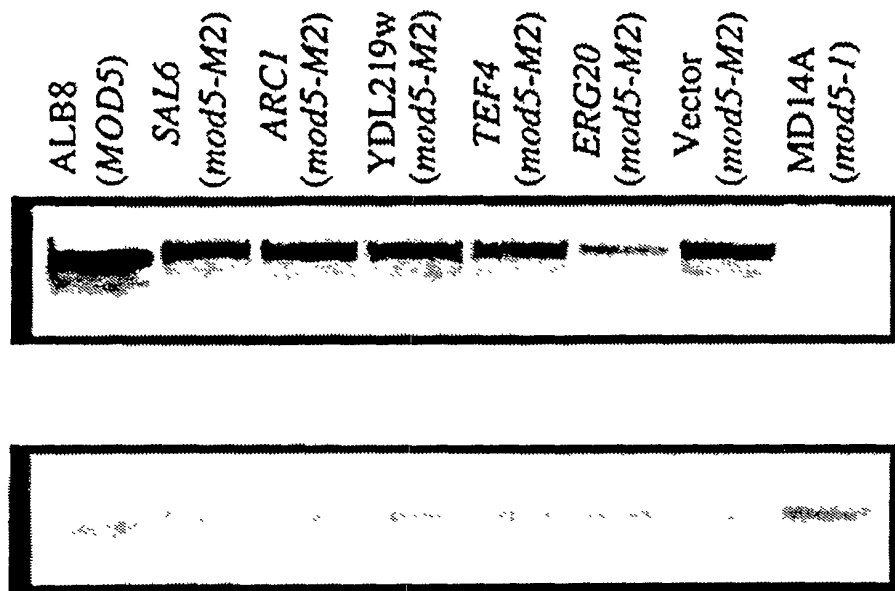


Fig. 1. The mevalonate pathway (11).

A.



B.

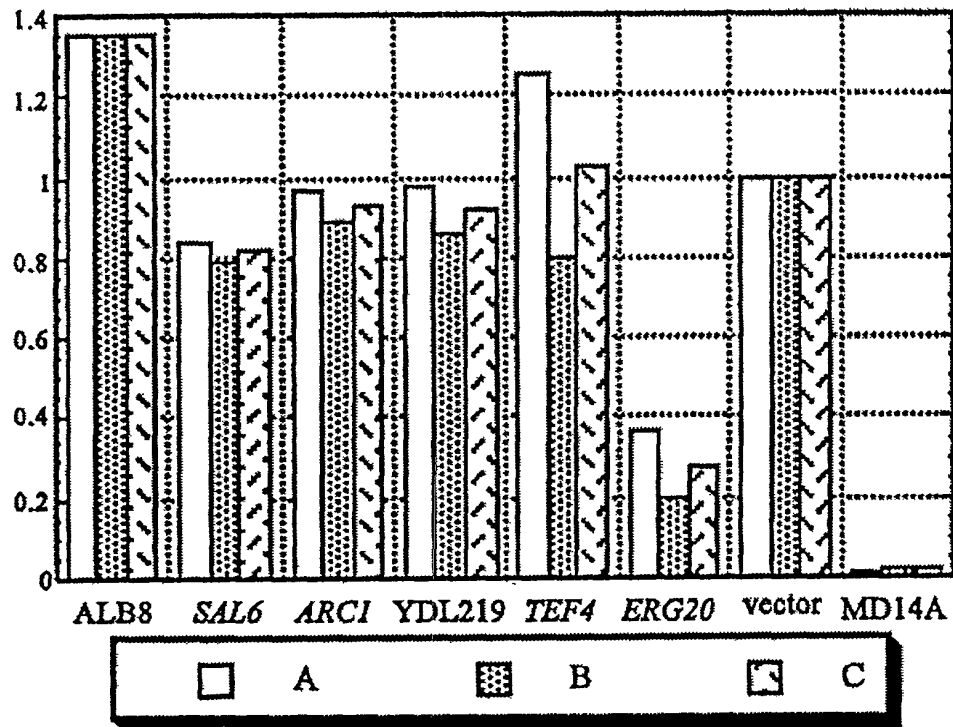
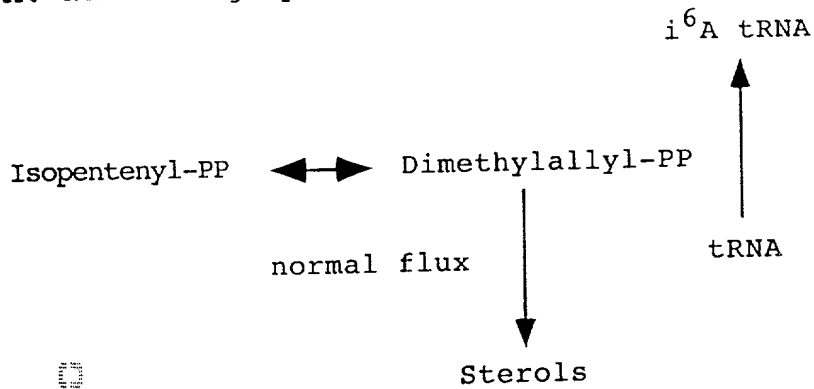


Fig. 2. The level of isopentenylated tRNA found in ALB1 overexpressing *ERG20* is reduced substantially. (A) Low-molecular-weight RNA was prepared from ALB1 (*mod5-M2*) with each of the candidate genes or vector alone, ALB8 (*MOD5*), or MD14A (*mod5-1*). The RNAs were resolved on polyacrylamide gels, transferred to membranes, and probed with anti-isopentenyl adenosine antibody (*Upper*) or radiolabeled oligonucleotide complementary to mature tRNA<sup>Tyr</sup> (*Lower*). (B) The levels of isopentenyl adenosine tRNA found in ALB1 with each of the candidate genes or vector only or in the strain ALB8 or MD14A were assessed by densitometric analysis of two immunoblots and expressed as a fraction of the level found in the "vector" control. A, membrane 1 values; B, membrane 2 values; C, average values.



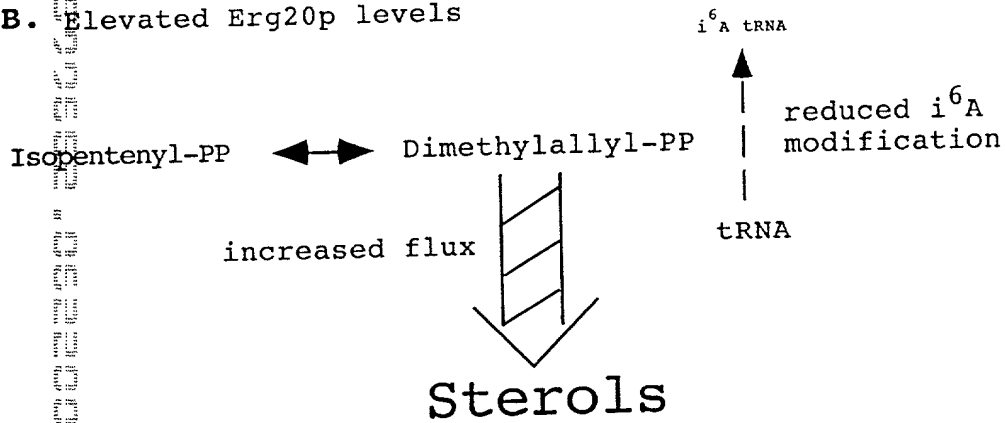
Phenotype of ALB1 cells

**A. Normal Erg20p levels**



white/light pink colonies; growth on -ade media; no growth on canavanine-containing media

**B. Elevated Erg20p levels**



pink/red colonies; poor growth on -ade media; able to grow on canavanine-containing media

**Fig. 3.** Model of competition between  $i^6$ A modification of tRNA and sterol biosynthesis.

NSII

M2           Snabi         ↓         ↓  
WT      TATACTCATATCTTTTGTAATAAGAACCAAAATCTTCATG CTA AAG GGA CCG CTT AAA GGT TGC TTA AAT ATG TCT AAA //GAA TGC ATG  
MET Leu Lys Gly Pro Leu Lys Gly Cys Leu Asn MET Ser Lys Glu Cys Met

M1                          HindIII  
AATCTTCAAG.CTT AAG GGA CC

SspI

M2           GGT TGC TTA AATATT TCT AA  
Gly Cys Leu Asn Ile Ser

## FIGURE 4

[illegible]

# Phenotype of Yeast

ALB1

T8-1D (YGmod5) M2 KR6

white/pink colonies	dark pink colonies
slow growth on -ade	poor growth on -ade
weak growth on canavanine-containing media	no growth on -lys
	growth on canavanine-containing media

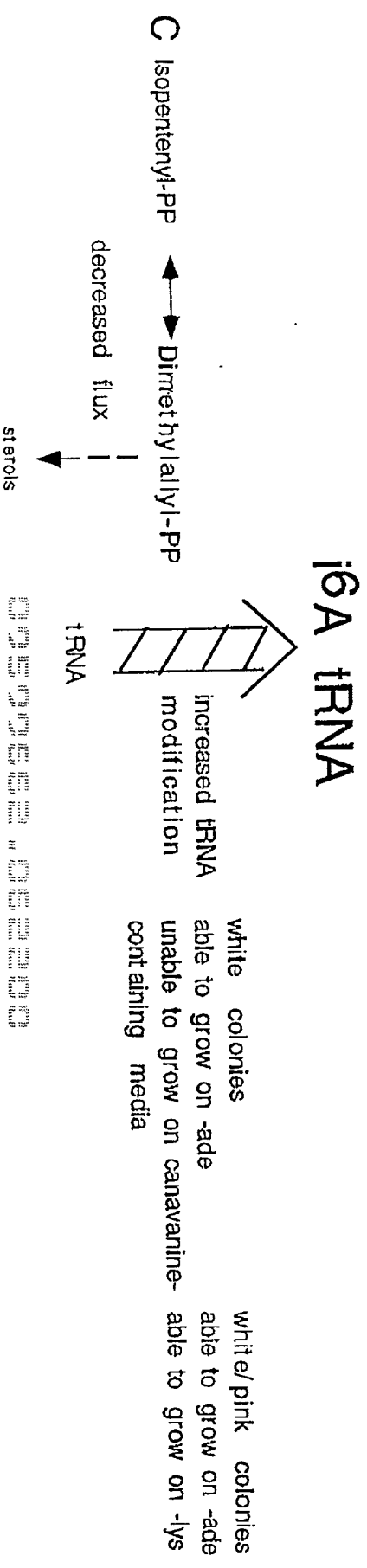
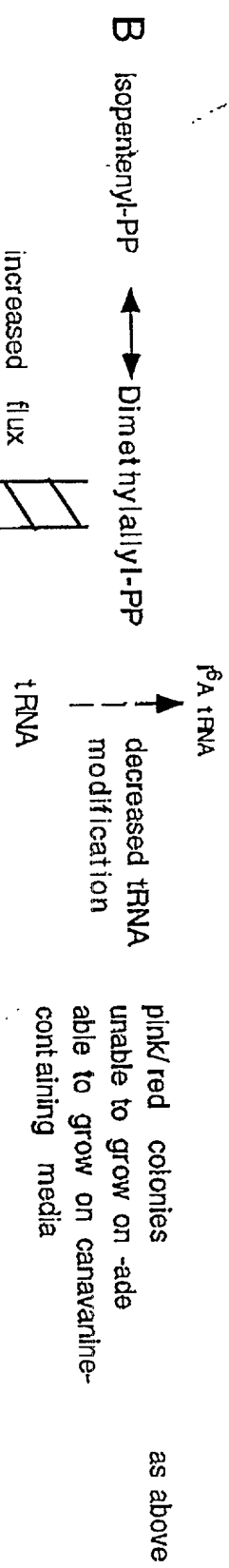
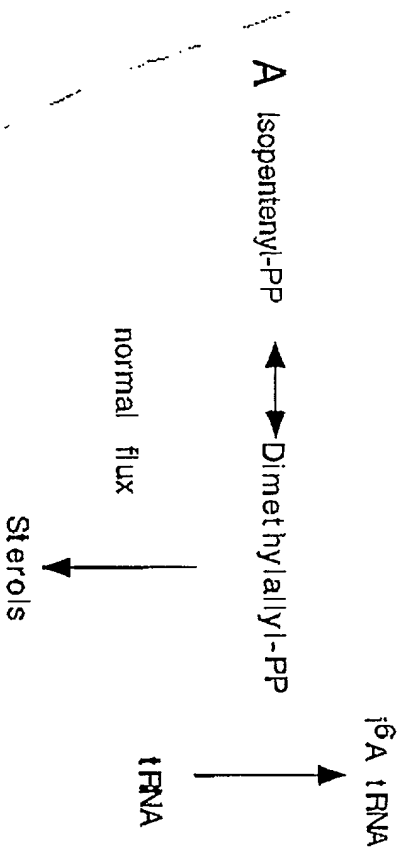
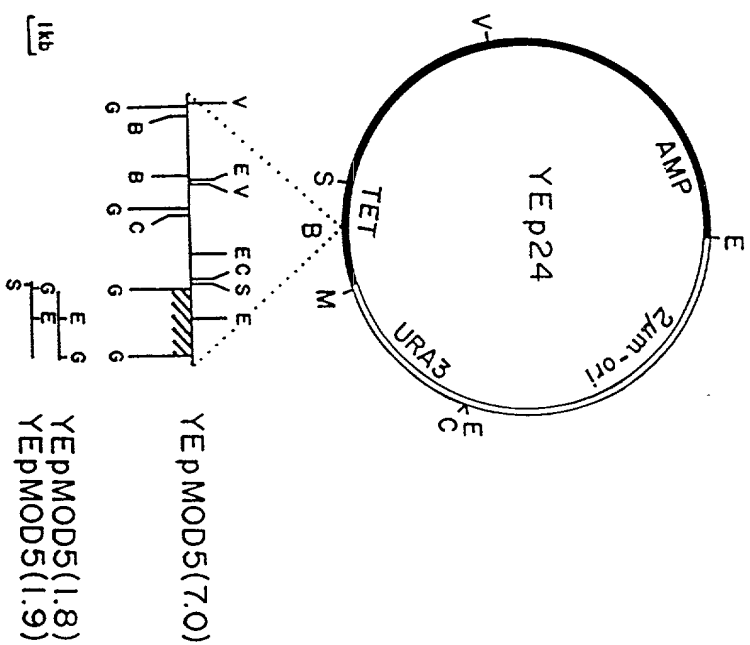


FIGURE 5



■, *E. coli* sequences of vector YE p24; □, insert of yeast genomic DNA; zzz: 1.8-kb *Bgl*II fragment used as a probe and fragment in YE pMOD5(1.8) and YCpMOD5 (1.8). B, *Bam*HI; C, *Cla*I; E, *Eco*RI; G, *Bgl*II; M, *Sma*I; S, *Sal*I; V, *Pvu*II.

# FIGURE 6

**PATENT**

Attorney Docket No.: PSU 99-2085

**DECLARATION FOR PATENT APPLICATION**

As the below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated next to my name. I believe I am the original, first and sole inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled **Compositions and Methods for the Quantification of Sterol Biosynthetic Flux**, the specification of which is attached hereto. I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

60/141,516	06/23/99	Pending
60/199,699	04/26/00	Pending
<i>Application Serial No</i>	<i>Filing Date</i>	<i>Patented, Pending or Abandoned</i>

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under § 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of Sole or First Inventor: **Anita K. Hopper**

Inventor's Signature: *Anita K. Hopper*  
 Residence: 8 Foxglove Drive, Hershey, Pennsylvania 17033  
 Post Office Address: 8 Foxglove Drive, Hershey, Pennsylvania 17033

Date: 06/20/00  
 Citizenship: United States of America

*Circle*

## DECLARATION FOR PATENT APPLICATION

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated next to my name. I believe I am an original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled **Compositions and Methods for the Quantification of Sterol Biosynthetic Flux**, the specification of which is attached hereto. I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

60/141,156  
60/199,699

06/23/00  
04/26/00

Pending  
Pending

Application Serial No.

Filing Date

Patented, Pending or Abandoned

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under § 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of Second Joint Inventor: Nancy C. Martin

Inventor's Signature:

Nancy C. Martin

Date:

6/20/00

Residence: Jouett Hall, 2301 S. Third Street, Louisville, Kentucky 40208

Citizenship: United States of America

Post Office Address: Jouett Hall, 2301 S. Third Street, Louisville, Kentucky 40208

13 River Hill Rd  
Louisville  
Ky 40207

## DECLARATION FOR PATENT APPLICATION

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated next to my name. I believe I am an original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled **Compositions and Methods for the Quantification of Sterol Biosynthetic Flux**, the specification of which is attached hereto. I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

60/141,156	06/23/99	Pending
60/199,699	04/26/00	Pending
<hr/> Application Serial No.	<hr/> Filing Date	<hr/> Patented, Pending or Abandoned

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under § 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of Third Joint Inventor: **Ann Benko**

Inventor's Signature: Ann Benko  
Residence: 147 Woodridge Drive, Palmyra, Pennsylvania 17078  
Post Office Address: 147 Woodridge Drive, Palmyra, Pennsylvania 17078

Date: 6/20/00  
Citizenship: United States of America

## DECLARATION FOR PATENT APPLICATION

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated next to my name. I believe I am an original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled **Compositions and Methods for the Quantification of Sterol Biosynthetic Flux**, the specification of which is attached hereto. I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

60/141,516	06/23/99	Pending
60/199,699	04/26/00	Pending
<i>Application Serial No.</i>	<i>Filing Date</i>	<i>Patented, Pending or Abandoned</i>

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under § 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of Fourth Joint Inventor: Gabriela Vaduva

Inventor's Signature: Gabriela Vaduva

Date: June 20, 2000

Residence: 10317 Sannois Drive, Apt. 5, St. Louis, Missouri 63146

Citizenship: Romania

Post Office Address: 10317 Sannois Drive, Apt. 5, St. Louis, Missouri 63146